

(19) World Intellectual Property Organization
International Bureau



(43) International Publication Date
14 March 2002 (14.03.2002)

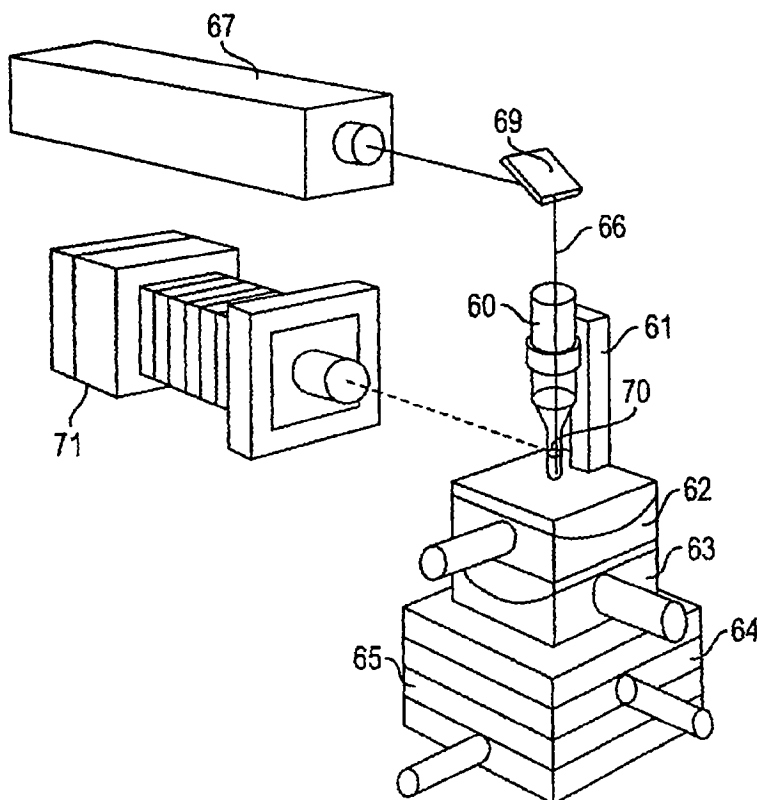
PCT

(10) International Publication Number
WO 02/21108 A3

- (51) International Patent Classification⁷: **G01N 21/51**, 21/64 (72) Inventors; and
(75) Inventors/Applicants (for US only): **ANDERSON, Norman, G.** [US/US]; 10916 Wickshire Way, Rockville, MD 20852 (US). **ANDERSON, N., Leigh** [US/US]; 1759 Willard Street, NW, Washington, DC 20009 (US).
- (21) International Application Number: **PCT/US01/27529**
- (22) International Filing Date:
6 September 2001 (06.09.2001) (74) Agents: **IHNEN, Jeffrey, L.** et al.; Rothwell, Figg, Ernst & Manbeck, P.C., 555 13th Street, N.W., Suite 701-E, Washington, DC 20004 (US).
- (25) Filing Language: **English**
- (26) Publication Language: **English** (81) Designated States (national): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, PH, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW.
- (30) Priority Data:
09/657,531 8 September 2000 (08.09.2000) US
- (71) Applicant (for all designated States except US): **LARGE SCALE PROTEOMICS CORPORATION** [US/US]; 9620 Medical Center Drive, Rockville, MD 20850 (US).

[Continued on next page]

(54) Title: METHOD FOR DETECTING MOLECULES OR CHEMICAL REACTIONS BY DETERMINING VARIATION OF CONDUCTANCE



(57) Abstract: A method for separating microorganisms, especially infectious agents, from a mixture by two dimensional centrifugation on the basis of sedimentation rate and isopycnic banding density, for sedimenting such microorganisms through zones of immobilized reagents to which they are resistant, for detecting banded particles by light scatter or fluorescence using nucleic acid specific dyes, and for recovering the banded particles in very small volumes for characterization by mass spectrometry of viral protein subunits and intact viral particles, and by fluorescence flow cytometric determination of both nucleic acid mass and the masses of fragments produced by restriction enzymes. The method is based on the discovery that individual microorganisms, such as bacterial and viral species, are each physically relatively homogeneous, and are distinguishable in their biophysical properties from other biological particles, and from non-biological particles found in nature.



(84) **Designated States (regional):** ARIPO patent (GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, TR), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG).

— *before the expiration of the time limit for amending the claims and to be republished in the event of receipt of amendments*

(88) **Date of publication of the international search report:**

23 May 2002

Published:

— *with international search report*

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

DETECTION AND CHARACTERIZATION OF MICROORGANISMS

CROSS-REFERENCE TO RELATED APPLICATION

5 The present application is a continuation-in-part of application Serial No. 09/265,541 filed 9 March 1999 and is related to U.S. provisional patent application Serial No. 60/077,472, filed on 10 March 1998, both of which are incorporated herein by reference.

10 This invention was made with Government support under an SBIR grant from NIH, Grant No. 1 R43 AI41819-01/02. The United States government may have certain rights in the invention.

BACKGROUND OF THE INVENTION

15 The present invention relates to the field of separating and identifying microorganisms, particularly infectious agents, using two-dimensional centrifugation and exposure to chemical and enzymatic agents, combined with detection in density gradients based on light scatter or fluorescence, counting by fluorescence flow cytometry, and characterization of intact virions, bacteria, proteins and nucleic acids by mass spectrometry, flow cytometry and epifluorescence microscopy.

20 The publications and other materials used herein to illuminate the background of the invention or provide additional details respecting the practice, are incorporated by reference, and for convenience are respectively grouped in the appended List of References. Patents referenced herein are also incorporated by reference.

25 In the prior art, diagnosis of viral and bacterial infections has been done by culturing the causal agents in suitable media or in tissue culture to obtain sufficient particles for analysis, followed by identification based on which conditions support growth, on reaction to specific antibodies, or based on nucleic acid hybridization (Gao and Moore, 1996). Biological growth can be omitted when the polymerase chain reaction (PCR) is used to amplify DNA, however, PCR requires sequence-specific primers, and is thus limited to known or suspected agents (Bai et al., 1997). For all these methods, considerable time is required, and the methods are useful for agents
30 whose properties are known or suspected. Existing methods do not provide means for rapidly isolating and characterizing new infectious agents. Hundreds of infectious agents are known, and it is infeasible to have available reagents for an appreciable fraction of them.

Techniques for recovering infectious agents from blood, urine, and tissues have been previously developed based on centrifugation or filtration, but have not been widely used clinically (Anderson et al., 1966; Anderson et al., 1967). The highest resolution methods use rate zonal centrifugation to separate fractions based on sedimentation rate (measured in Svedberg units, S) and isopycnic banding density (measured in grams per mL or ρ). S- ρ separations have been used to isolate virus particles in a high state of purity from rat liver homogenates, and have been used to isolate the equivalent of approximately 20 virions per cell (Anderson et al., 1966). In these studies, virus particles were detected by light scattering and visualized by electron microscopy. The separations required complex special equipment not generally available, one or more days of effort, and they did not provide a definitive identification of the bacterial or viral species separated.

It is important to show that candidate infectious particles isolated by centrifugal methods actually contain nucleic acids. DNA and RNA in both active and fixed bacterial and viral particles have been stained with fluorescent dyes specific to nucleic acids, and observed and counted by fluorescent microscopy and flow cytometry. Many dyes are now known which exhibit little fluorescence in the free state, but become highly fluorescent when bound to nucleic acids. Some bind differentially to DNA or RNA or to different specific regions, and some show different emission spectra depending on whether bound to DNA or RNA. In this disclosure, dyes referred to are fluorescent dyes. By differential fluorescence spectroscopy ssDNA, dsDNA and RNA may be distinguished. See, Haugland, 1996; Mayor and Diwan, 1961; Mayor, 1961; Hobbie et al., 1977; Zimmerman, 1977; Perter and Feig, 1980; Paul, 1982; Suttle, 1993; Hirons et al., 1994; Hennes and Suttle, 1995; Hennes et al., 1995.

Isolated nucleic acid molecules of the dimensions found in bacteria and viruses have been counted and their mass estimated using fluorescence flow cytometry for molecules in solution, and epifluorescence microscopy of immobilized molecules (Hennes and Suttle, 1995, Goodwin et al., 1993). In both instances, the size of fragments produced by restriction enzymes can be estimated, and the molecules identified by reference to a database listing the sizes of fragments of known DNA molecules produced by different restriction enzymes (Hammond et al., U.S. Patent No. 5,558,998; Jing et al., 1998).

Using specific fluorescently-labeled antibodies, specific identifications may also be made. These studies are time consuming, and require batteries of specific antibodies, together with epifluorescent microscopy or fluorimeters.

Matrix-Assisted Laser-Desorption-Ionization Time-of-Flight Mass Spectrometry (MALDI-TOF-MS) currently allows precise measurements of the masses of proteins having molecular weights of over 50,000 daltons. Individual virion proteins have been previously studied by mass spectrometry (Siuzdak, 1998); however, resolution of complete sets of viral subunits from clinically relevant preparations of intact viruses, and the demonstration that precise measurements could be made of their individual masses, have not been previously reported. While single protein mass measurements can reliably identify many proteins, when a set of proteins from a virus or bacterial cell are known, detection of such a set provides more definitive identification. Methods are currently also being developed which allow partial sequencing of proteins or enzymatically produced peptide fragments and thus further increase the reliability of identifications. For MALDI-TOF-MS currently used methods require a picomole or more of protein, while electrospray mass spectrometry currently requires 5-10 femtomoles. The detection limits with mass spectrometry, especially MALDI, depend on getting a sample concentrated and on to a very small target area. Sensitivity will increase as ultramicro methods for concentrating and transferring ever smaller-volume samples are developed. See, Claydon et al., 1996; Fenselau, 1994; Krishnanurthy et al., 1996; Loo et al., 1997; Lennon and Walsh, 1997; Shevchenko et al., 1996; Holland et al., 1996; Liang et al., 1996.

Centrifugal methods for concentrating particles from large into small volumes have been in use for decades. Using microbanding centrifuge tubes which have a large cylindrical volume and cross section which tapers gradually in a centrifugal direction down to a small tubular section, particles may be concentrated or banded in a density gradient restricted to the narrow tubular bottom of the tube, or may be pelleted. The basic designs of such tubes are well known by those skilled in the arts. See, Tinkler and Challenger, 1917; Cross, 1928; ASTM Committee D-2, 1951; Davis and Outenreath, U.S. Patent No. 4,624,835; Kimura, U.S. Patent No. 4,861,477; Levine et al., U.S. Patent No. 5,342,790; Saunders et al., U.S. Patent No. 5,422,018; Saunders, U.S. Patent No. 5,489,396. The original tubes of this type were called Sutherland bulbs and were used to determine the water content of petroleum (The Chemistry of Petroleum and Its Substitutes, 1917, ASTM Tentative Method of Test for Water and Sediment by Means of Centrifuge, ASTM Designation: D 96-50T, 1947). Slight modifications of the basic design are described in U.S. Patent Nos. 4,106,907; 4,624,835; 4,861,477; 5,422,018; and 5,489,396. Such tubes have been made of glass or plastic materials, and the use of water or other fluids to support glass or plastic centrifuge tubes in metal centrifuge shields has long been well known in the art. However, centrifuge tubes disclosed

in the prior art which include a shape similar to that of the microbanding centrifuge tubes of the instant invention could not withstand the centrifugal forces required to band viral particles in gradients. Conventional centrifuge tubes, or tubes derivative from the Sutherland design have been used for density gradient separations, and for separations in which wax or plastic barriers are used which position themselves between regions of different density to allow recovery of these fractions without mixing. There has been no previous discussion of barriers which prevent mixing of step gradient components at rest, but which barriers are centrifuged away from the gradient during rotation. Nor have tube closures for high-speed thin-walled swinging-bucket centrifuge tubes been described, whose exterior surfaces can be disinfected after the tubes are loaded.

The efficient stabilization of very shallow density gradients in centrifugal fields is well known, and is utilized in analytical ultracentrifugation to cause a sample layer to flow rapidly to the centripetal surface of a gradient without mixing using a synthetic boundary cell (Anderson, U.S. Patent No. 3,519,400). Hence, light physical barrier disks between step gradient components can be moved away from the gradient by centrifugal force without appreciably disturbing the gradient, provided that they are made of porous, woven or sintered materials having a physical density less than that of the sample layer, such as polyethylene or polypropylene.

Many authors have noted that viruses and bacteria are often resistant to the actions of detergents and enzymes which will digest or dissolve contaminating particles of biological origin, and efforts have been made to classify infectious agents on the basis of their differential sensitivities. These differences have not previously and conveniently been incorporated in a method for detecting and quantifying infectious agents. See, Gessler et al., 1956; Theiler, 1957; Epstein and Hold, 1958; Kovacs, 1962; Planterose et al., 1962; Gard and Maaloe, 1959. Density differences between different species of virus and bacteria are well known, but have not been previously exploited for purposes of identification.

Infectious particles exhibit a wide range of isopycnic banding densities ranging from approximately 1.17 g/ml to 1.55 g/ml, depending on the type of nucleic acid present, and the ratios between the amount of nucleic acid, protein, carbohydrate, and lipid present. While such banding density differences are well known, no attempt has been previously made to systematically measure them and use the data to classify infectious agents.

The present invention is directed to an integrated system for concentrating, detecting and characterizing infectious agents using separations based on sedimentation rate and banding density, spectral analysis of emitted fluorescent light to distinguish DNA from RNA, differentiation of viral

and bacterial particles from other particles by sedimentation through zones of solubilizing enzymes or reagents, determination of the isopycnic banding densities of infectious particles by reference to the positions of synthetic density standardization particles, particle detection using fluorescent dyes for DNA or RNA, further concentration of banded particles by pelleting, transfer of concentrated particles to mass spectrometer targets for protein mass determination and analysis, counting of concentrated particles by epifluorescent microscopy and fluorescence flow cytometry, and identification of bacterial or viral nucleic acids by restriction fragment length polymorphism analysis using either immobilized nucleic acid molecules, or ultrasensitive fluorescence flow cytometry. These methods are especially useful in characterizing biological samples which have low titres of virus and which contain viruses which are not culturable.

Furthermore, all current methods used to detect and characterize infectious agents, including use of fluorescent antibodies, detection of agent-associated enzymes, culture to increase agent mass, PCR amplification, restriction fragment length polymorphism analysis, hybridization to probes immobilized on chips, histochemical analysis, and all forms of microscopy including electron microscopy, are vastly improved by preconcentration of the microorganisms using the methods of the present invention.

The present invention is further directed to a method and apparatus for measuring light scatter from a microbanding zone.

These techniques have not previously been assembled into one operational system capable of routine field, hospital, and clinical laboratory use. The present application describes innovations and inventions which make such a system feasible. For work with potentially lethal agents, the system will be assembled in containment, and at least partially automated.

SUMMARY OF THE INVENTION

The overall objective of this invention is to develop a physical system for rapidly identifying infectious disease agents without growing them, and for discovering new infectious agents. The process is based on the thesis that infectious agents constitute a unique group of particles which can be isolated by physical and chemical means from other natural particles and identified by their physical parameters using centrifugal means, light scatter, fluorescence, and mass spectrometry. The system will allow rapid clinical distinction between viral and bacterial infections, identification of specific agents with the aim of providing specific therapy, and the rapid discovery of new infectious agents. In addition the system will make it feasible to develop and test new antibiotics

and antiviral agents in man by measuring the effects of these agents on bacterial and viral loads. At present the development of new antiviral drugs is severely hindered by inability to define populations of individuals in the early stages of infection who might benefit from treatment.

In accordance with the present invention, an ultracentrifuge tube is provided which comprises upper, middle and lower regions of successively smaller diameters. In one embodiment, the tube has an upper region for receiving a sample, a funnel-shaped middle region and a lower narrow tubular microbanding region. The diameter of the lower region may be 0.25 inch or less, preferably 0.1 inch or less, more preferably 0.1 to 0.08 inch, and most preferably 0.08 to 0.039 inch. Smaller diameter microbanding regions are feasible and are within the province of this disclosure. The length of the lower region is typically between 5% to 25% of the length of the tube. In one aspect, the tube may also include a seal with a central opening which can be plugged and unplugged.

In accordance with the present invention, a bucket is provided for holding an ultracentrifuge tube. The bucket comprises upper and lower regions which may be of successively smaller diameters, may have inserts to successively decrease the inner diameter, or may be of uniform internal diameter, and may further comprise a third region which attaches the bucket to a rotor.

Further in accordance with the present invention, a method is provided for concentrating microorganisms. As used herein, the term "microorganisms" is intended to include viruses, mycoplasmas, rickettsia, yeast and bacteria. The method comprises ultracentrifugation of a sample containing the microorganism in an ultracentrifuge tube described herein. The ultracentrifugation may include the formation of density gradients and/or the staining of the microorganism(s). In one aspect, the staining can be used to distinguish the DNA or RNA content of a virus. The banding of the microorganisms upon ultracentrifugation can be used to identify the microorganisms.

In a further aspect of the invention, the concentrated microorganisms are further characterized by conventional techniques such as mass spectrometry, flow cytometry, optical mapping, isopycnic banding densities, light scattering, fluorescence, restriction enzyme analysis, genome size, enzymatic or chemical resistance/susceptibility, immunochemistry and the like. In another aspect of the invention, the amount or titre of the microorganisms can be determined.

In accordance with the present invention, a system is provided for measuring light scatter and fluorescence from a sample in a centrifuge tube. In one embodiment, the system includes a centrifuge tube, a light source, such as a laser, and a detector to detect light passing through the sample or emitted or scattered from the sample upon light passing through it. Optical filters select and separate the exciting and emitted or scattered wavelengths of light.

The internal surfaces of the tubes and especially the funnel portion must be very smooth in order to prevent small virus particles from being retarded by surface irregularities and, in addition, the surfaces must be treated so that infectious particles are not adsorbed. Polishing of plastic surfaces is done by brief exposure to a solvent vapor. For example, polycarbonate is polished by
5 brief exposure to heated methylene chloride gas. Plastic surfaces are modified to prevent adsorption of infectious particles by exposure to dilute solutions of proteins such as bovine serum albumin or gelatin, or to charged polymers such as heparin or derivatives of heparin. Both of these procedures are well known to those practiced in the arts.

Further in accordance with the present invention, a system is provided for counting particles
10 concentrated in a small volume. The system includes a container in which the particles are concentrated, a capillary tube, two pumps, means for moving the container relative to the capillary tube, a flow cell, a light source and detector. Alternatively, fluid may be moved by gas pressure instead of pumps.

BRIEF DESCRIPTION OF THE DRAWINGS

15

Figure 1 shows an S-rho plot for a typical tissue and for representative viruses.

Figures 2A-2C are a diagrammatic representation of one embodiment of a centrifugal microbanding tube and its use.

Figures 3A-3G show alternative embodiments of microbanding tubes and use in a rotor (3F).
20

Figures 4A and 4B illustrate a centrifuge swinging bucket design that allows higher speed fractionation of large sample volumes.

Figure 5 illustrates a complete system including vertical monochromatic laser illumination, goniometer and X-Y stage for supporting and positioning microbanding tube, microbanding tube with banded viruses particles, and camera system.

Figure 6 illustrates a complete system including interference filter light sources, light pipe illumination, digital data acquisition, and CRT data presentation.
25

Figure 7 illustrates a method for recovering banded virus particles using a micropipette, and counting them by flow cytometry.

Figures 8A-8C illustrate details of band recovery.

Figures 9A-9F illustrate one embodiment of a closure for swinging bucket rotor centrifuge tubes and detection of a sample with respect to the tubes.
30

Figure 10 illustrates a complete system that is similar to the system depicted in Figure 5, the system in Figure 10 used to measure light scatter in accordance with the present invention.

Figure 11 is a computer generated graph indicating the intensity of measured light scatter from a microbanding zone in a centrifuge tube in accordance with the present invention.

5 Figure 12 is an image from the camera of the system depicted in Figure 10 showing the light scatter from the microbanding zone in the centrifuge tube in accordance with the present invention.

Figure 13 is a computer generated graph indicating the intensity of measured fluorescence from the microbanding zone in the centrifuge tube in accordance with the present invention.

10 Figure 14 is an image from the camera of the system depicted in Figure 10 showing the fluorescence from the microbanding zone in the centrifuge tube in accordance with the present invention.

Figure 15 is a schematic drawing showing the system depicted in Figure 10, with elements removed to provide greater clarity and show the orientation of the polarized light from the laser in accordance with the present invention.

15 Figure 16 is another schematic drawing showing the system depicted in Figure 10, with elements removed to provide greater clarity, and showing moveable filters that enable the system to measure both light scatter and fluorescence in accordance with the present invention.

DETAILED DESCRIPTION OF THE INVENTION

20 The invention is directed to methods of identifying and measuring the presence of microbial agents such as bacteria and viruses in biological samples. The methods include centrifugation steps to purify the microbial agents in a very small volume. The agents are then assayed by means such as isopycnic banding density, light scatter detection, fluorescence or mass spectrometry.

25 It is an object of this invention to develop integrated systems and methods in which suspensions containing microorganisms, including infectious agents, are stained with one or more fluorescent dyes, in which a step or continuous gradient is automatically formed during centrifugation, in which the microorganisms are centrifuged away from the stain-containing suspending medium and are washed free of external stain, are concentrated in a gradient of very small cross section, separated according to their isopycnic banding densities, their banding densities
30 determined, and the microorganisms detected by fluorescence.

It is a further object of this invention to concentrate microorganisms, including infectious agents, into microbands by a factor of 1-5,000.

It is a further object of this invention to expose the microorganisms, such as infectious agents, to reagents including detergents, surfactants, enzymes, or organic solvents contained in distinct zones in a density gradient to dissolve or disassemble contaminating particles to prevent them from banding with the microorganisms, and to separate stained particles from the free stain of the initial sample volume.

It is a further object of this invention to use one or more dyes which bind differentially to RNA, single stranded DNA or double stranded DNA to allow these to be distinguished by their fluorescent spectra.

It is a further object of this invention to provide for the concentration of banded microorganisms, for example infectious agents, by resuspending the banding gradient, which is typically 0.04 mL, to approximately 4 mL in water or a very dilute buffer, and pelleting the microorganisms one or more times to provide a concentrated pellet free of gradient materials for mass spectrometric analysis, for counting by epifluorescent microscopy or by flow cytometry.

It is a further object of this invention to provide means for the diagnosis of infectious diseases which minimize exposure of laboratory personnel to infectious agents.

It is an additional object of this invention to provide means for preparing nucleic acids from small quantities of microorganisms, including infectious agents, to determine the masses of intact nucleic acid molecules, and for characterization of fragments produced by restriction enzymes using either flow cytometry or epifluorescence microscopy.

It is an additional object of this invention to determine the banding densities of the microorganisms, such as infectious agents, accurately by reference to the positions of calibrated particles added to the gradients.

For ease of description, the invention will be described with reference to viruses as the microorganisms. It will be understood that the invention is also applicable to other microorganisms, including mycoplasmas, yeast and bacteria. The invention is particularly suited for the identification of infectious agents, and will be described in this context.

Figure 1 is a graph depicting the sedimentation coefficients and isopycnic banding densities of subcellular organelles and viruses to illustrate the concept of the "Virus Window" (Anderson, 1966). It is evident that viruses have a relatively narrow range of sedimentation coefficients and banding densities and may be isolated from a tissue homogenate or from blood in a high state of purity using high resolution S- ρ separation systems. For a complete description of high resolution S- ρ centrifugal methods and of centrifuge development for virus isolation, refer to National Cancer

Institute Monograph 21, The Development of Zonal Centrifuges and Ancillary Systems for Tissue Fractionation and Analysis, U.S. Department of Health, Education and Welfare, Public Health Service, 1966. This work describes S- ρ separation theory and systems, and the use of colored plastic beads of graded densities as density markers.

5 In practice, a blood sample or tissue homogenate is centrifuged to sediment all particles having sedimentation rates higher than that of the particle or particles to be analyzed. For viruses, that means particles of circa 10,000 S and above are discarded. The supernatant after such a separation is then used as the sample for second dimension isopycnic banding separations carried out in microbanding tubes as described here, using centrifugation conditions which will sediment and isopycnicly band all known infectious particles.

10 One picomole of virus would contain 6.022×10^{11} viral particles, while 6×10^9 virions would contain 1 picomole of a viral coat protein present in 100 copies per virion. Quantitative polymerase chain reaction (PCR) has been used to demonstrate that in many infectious diseases $> 10^8$ virus particles are present per mL of plasma or serum. Hence, if the virions from a 5-10 mL biological sample containing 10^8 virions/mL are concentrated to a microliter or two, and then applied to a very small target area, individual viral proteins can be detected using MALDI-TOF-MS (Krishnanurthy et al., 1996; Holland et al., 1996). Using electrospray techniques, samples containing 10^6 virus particles/mL can be detected, while with flow cytometry and immobilized DNA epifluorescence microscopy, even fewer particles are required (Hara et al., 1991; Hennes and Suttle 1995). The application of these methods to bacteria may require a preseparation of proteins to reduce the complexity of the sample. In mass spectrometry, detection has been by charged ion detection, and the limitations of such detection have set the upper limits to the size of proteins and nucleic acids that can be detected. Mass spectrometric methods have now been described which allow masses of biological particles above 100,000 daltons to be measured.

25 In order to work with such levels of virus, the virus must be concentrated into a very small volume. This concentration is accomplished during the second dimension of centrifugation (the isopycnic banding step) by banding the virus using a centrifuge tube specially designed to concentrate the virus into a microband after passage through gradient layers that wash the particles and expose them to selected reagents. An example of such a microbanding centrifuge tube is shown in Figures 2A-2C. Figure 2A illustrates diagrammatically a hollow transparent centrifuge tube 1 with an upper sample volume 2, grading into a serrated funnel region 3 having successively tapered and parallel-wall sections 4-6, constricting down to a narrow tubular microbanding region 7. The

serrated funnel region 3 is an improvement over centrifuge tubes which simply taper from top to bottom without including a serrated region. By serrations is meant, for example, concentric rings or edges or lips. These rings, edges or lips are preferably continuous around the inner diameter of the centrifuge tube, but this is not required. For example, three projections from the inner wall of the centrifuge tube spaced equally around the diameter could be used to hold a disk in place. The term serrations is meant to include such possibilities but does not include a straight tapering with no rings, edges, lips or projections on the inner surface of the centrifuge tube. The serrations can be used as rests onto which disks can be placed to separate two or more layers of liquid. Although disks can be placed into tubes which simply taper without serrations, the disks in such tapered tubes can be easily tipped up on one edge by pushing down on the opposing edge. This would cause a premature mixing of the layers which are to be separated by the disks. The serrated region allows disks to lie flat and prevents the disks from being accidentally tipped up. As an example, the centrifuge tube may be 3.45 inches from top to bottom, have an outer diameter at the top of 0.562 inch, and have an inner diameter in the bottom microbanding region 7 of 0.064 inch. Such a tube is suitable for use in an SW41 Ti (Beckman) rotor. The inside surface of the tube is preferably polished using conventional techniques, including vapor polishing, so that the virus particles do not stick to the wall of the tube. Additionally, the internal surfaces of the tubes may be coated with a protein or polymer to prevent particle adhesion, as is well known in the art.

Figure 2B illustrates how the tube is loaded at rest with a series of fluids of decreasing physical density. The tube shown comprises a series of serrations onto which can be laid disks to separate one layer of fluid from the next layer of fluid. Liquid 8 is denser than any particle to be recovered, and is used to partially fill the microbanding region 7. When a less dense fluid 9 is pipetted in with a micropipet an air bubble 10 (wherein by air is meant atmospheric air or another gas) may be left to keep the fluids 8 and 9 separate. Similarly when the first overlay fluid 11 is introduced, air bubble 12 may be left in place, thus keeping the three liquids separate until centrifugation is commenced. A tube with an inner diameter of 0.064 inches in microbanding region 7 is suitable for allowing an air bubble to be left in place to separate two layers of liquid. Alternatively, the air bubbles may be left out and the fluids allowed to diffuse together to create a density gradient. Fluid 11 is covered with a light porous plastic disk 13, preferably of sintered polyethylene or polypropylene, which fits in place in the first serration. A fluid 14, less dense than fluid 11, which may contain one or more reagents, is then introduced, and covered with disk 15, followed by even less dense liquid 16 which is covered with disk 17. The entire system is stable

until centrifuged. Before use the sample layer **18**, which has a density less than that of fluid **16** is then added up to level **19**. The tubes are then centrifuged at high speed in metal shields, typically with water or other liquid added to the shields. In addition, the tubes may be supported by fitting adapters which fill the space between the tubes and the shields, and water may be added to fill any spaces between the tubes, adapters and shields to provide additional support. Optionally the tubes may be capped (as shown in Figures 9A-9F, described in further detail below), to minimize the chances of operator infection.

Figure 2C illustrates diagrammatically a tube after centrifugation. The porous separation disks **13**, **15**, and **17** have risen to the top of the tube, and sample layer **18** is cleared of virus, and the original step gradient has changed, by diffusion, into one of a series of shallow gradients. In addition, gas bubbles **10** and **12** have also moved centripetally, and fluids **8** and **9** have come into contact to form a steep gradient by diffusion. As centrifugation proceeds, the slope of this gradient diminishes, producing a banding gradient of a width suitable for banding the infectious agents. For cesium chloride gradients, the densities typically range from 1.18 to 1.55 g/ml. These gradient steps may not only contain reagents to dissolve non-viral particles, but also serve to wash excess fluorescent dye away from the particles. For example, various detergents or enzymes such as proteases may be added either to the sample layer **18** or to other layers such as **14** or **16**. Fluorescent dyes may also be present in these regions. The free dye will not enter the lower, more dense regions in which the virus bands and therefore the centrifugation will purify the viruses from all of the reagents which may be present in the upper, less dense layers. After centrifugation, the microbanding region of the tube contains the upper portion of the banding gradient **27**, banded virus **28** (including any dye bound to the virus or viral nucleic acid) and lower dense portion of the banding gradient **29**, and the gradient formed between them by diffusion.

Figures 3A-3G illustrate alternative embodiments of tubes useful for microbanding of viruses and bacteria and all have a serrated internal construction which allows one or more light barriers to be positioned and retained at rest. Tubes shown in Figures 3A-3D and 3G are designed to be centrifuged in swinging bucket rotors so that the tubes are horizontal during centrifugation and vertical at rest. The tube shown in Figure 3A is the more conventional design with a sample reservoir **31**, a serrated funnel region **32**, and a microbanding section **33**. The tube shown in Figure 3B is similar to that of Figure 3A, but it is supported in a centrifuge shield by a support insert **34** which may be of plastic or metal. The tubes shown in Figures 3B-3E fill a rotor chamber completely. The tube of Figure 3C has an opaque bottom section **35** which absorbs scattered light,

while that shown in Figure 3D has a bulbous section 36 at the bottom of the microbanding tube 37 to contain an excess volume of the fluid forming the dense end of the gradient, thus stabilizing the gradient. The tube shown in Figure 3E is designed to be centrifuged in an angle head rotor as shown in Figure 3F, and has a linearly continuous wall 38 along one side positioned in the rotor so that particles may readily slide down to microbanding region 40. The tube shown in Figure 3G illustrates how a very large microbanding tube may be fabricated.

Figures 4A-4B illustrate how the tube of Figure 3G may be centrifuged at higher speed than tubes having a constant radius from top to curved bottom. This is accomplished by using a metal, plastic or carbon fiber shield 45 which matches the dimensions of tube 46. The shield has a cap 47 and the shield or bucket swings on integral attachment 48, as is conventionally done in high speed swinging bucket rotors. Tip 49 of the shield is much smaller diameter than the upper section of the shield, has much less mass swinging at its maximum radius, and hence can reach much higher speeds than is the case with shields of uniform internal diameter. This makes possible isolation of trace amounts of virus from much larger volumes than would otherwise be the case. During centrifugation using rotor 50 driven by drive 51, shield and tube 52 assume a horizontal position as shown.

The microbanded viruses can be analyzed at this stage or they can be collected, diluted, and further processed. To analyze the microbanded viruses at this stage, they can be detected by a system as shown in Figure 5. For example, the isopycnic banding step or an earlier step may have included a fluorescent dye or fluorescent dyes within the solution with which the virus was mixed or through which the virus was centrifuged. Dyes are known with which intact viruses may be stained and which can distinguish between RNA, DNA, single stranded nucleic acid and double stranded nucleic acid thereby allowing one to detect the presence or absence of an infectious agent, and further to determine which type of virus one has purified. The apparatus of Figure 5 can be used to analyze these stained particles.

A scanning and detection system is illustrated schematically in Figure 5 where microbanding tube 60 is held in a vertical position on mount 61 supported by goniometers 62 and 63 which are in turn supported by X-Y movements 64 and 65 in such a manner as to align and center the microbanding section of tube 60 with respect to laser beam 66. Laser beam 66 is generated by a laser 67, which may be an argon ion laser producing coherent light at 458, 488, 496, 502, and 515 nm. The beam passes through an interference or other filter 68 to isolate one wavelength, and is reflected down into the microbanding tube by dichroic mirror 69. The fluorescent banded particle

zone **70** is photographed or electronically scanned by camera **71** through emission filter **72**. The entire system may be enclosed to eliminate stray light, and filters **68** and **72** may be replaced by filter wheels (not shown) to optimize detection using fluorescent dyes which absorb and emit at different wavelengths, or to distinguish ssDNA, dsDNA and RNA by differences in the spectra of emitted fluorescent light. Electronic shutters may be attached to the laser to minimize sample exposure to light and to the camera to control exposure. The goniometers and X-Y movements may also be motor driven and remotely controlled, and the entire system may be controlled by a computer (not shown).

Figure 6 illustrates a different version of the scanning system which can cover all of the visible spectrum and on into the near ultraviolet. Microbanding tube **80** is aligned in a fixed support between transparent intensity equilibrators **81** and **82** attached to light pipes **83** and **84** which are in turn attached to intensity equilibrator **85** illuminated through filter **86** by condensing lens **87** and light source **88**. Filter **86** is one of a set attached to filter wheel **89** indexed by motor **90**. The result is uniform illumination from two sides of one or more bands **91**, **92** and **93**. The image is captured through emission filter **94** by digital camera **95** and the image stored, processed and displayed by computer **96** on CRT **97**. Filter **94** may be replaced by a filter wheel identical to **89** and **90** so that, with both an excitation filter wheel and an emission filter wheel and a wide spectrum light source such as a xenon lamp or a halogen lamp, a wide variety of combination of exciting and emitting light may be chosen, which in turn makes possible use of a wide variety of fluorescent dyes. Both fluorescent light and light scatter at a chosen wavelength may be employed for particle detection. This arrangement facilitates distinction between ssDNA, dsDNA and RNA.

The processed image **98** may be displayed to show a picture of the tube and contained bands **99**, **100** and **101**. The amount of light from each band may be integrated and displayed as peaks **102**, **103**, and **104**, and in addition the integrated values may be displayed digitally (not shown). The entire system including shutters on the light source and camera (not shown), filter movement and positioning, and focusing of the camera may be digitally controlled by computer **96**.

Display bands **99** and **101**, representing centrifuge tube bands **91** and **93** may be fluorescent or non-fluorescent density marker beads of known density, and the virus band **92** represented by display band **103**. The banding density of the virus may be determined by interpolation from the positions of the density markers. When non-fluorescent density markers are used, these are detected by scattered light using identical filters at positions **86** and **94**. A second image using suitable and different filters is then captured which is comprised solely of fluorescent light. The two images are

electronically inter-compared and the physical density of the infectious agent determined by interpolation.

At this stage, the virus can be identified as being a DNA virus or an RNA virus, and if a DNA virus it can be determined whether it is single stranded or double stranded. Furthermore, the density of the virus can be determined. This data can be used to help identify the type of virus which has been purified. Nevertheless, it may be desirable or necessary to gather more data to fully determine what the exact virus is and also to determine the original viral titre.

Figure 7 illustrates diagrammatically counting of individual fluorescent particles recovered from a tube 110 containing zones of banded virus 111 and 112 after all fluid above the banding gradient has been removed and replaced. The tube is placed in a tube holder 113, and an overlay of deionized water or very dilute buffer 114 is introduced above the gradient supplied through tube 115 to replace the volume drawn up in the probe 117. The tube may be closed at the top by a plastic closure 116. The capillary probe 117 is held stationary, and the microbanding tube 110 is slowly raised under it. The tube holder 113 is part of a precision drive mechanism 118 and associated stepping motor 119 that moves the tube holder vertically at a very slow and controllable rate. A slow steady stream of fluid is drawn into constriction 120 which is centered in sheath stream 121 provided by pump 122. The result is a constant flow of fluid through flow cell 123 with a fine virus containing stream in the center, elongated and extended by the flowing sheath. A second pump 124 withdraws fluid upward at a constant rate from the flow cell, which rate is greater than the rate at which piston pump 122 injects fluid into sheath 121. The difference in the rates of pumps 124 and 122 is made up by the fluid coming through capillary probe 117. The fluid coming through capillary probe 117 is a mixture of virus plus fluid from the overlay which is introduced via tube 115.

The flow cell 123 is illuminated by laser beam 125 produced by laser 126, that passes through exciting filter 127. Emitted light is isolated by emission filter 128 and detected by a photomultiplier 129. The output from the photomultiplier 129 is integrated at intervals by computer 130, and the integrated signal vs. time is displayed on CRT 131. When two viral bands are present, two peaks such as 132 and 133, are displayed. Depending on the number of fluorescent particles present, the signal generated from a band may be integrated into a peak, or, if the suspension is sufficiently dilute, the particles may be counted individually, the values binned, and the integrated results displayed.

In order to count the particles as just described, it is necessary that the virus particles are greatly diluted as they pass through flow cell 123. Figure 8 illustrates diagrammatically how the problem of making an initial dilution of a very small-volume virus band for counting individual particles is accomplished. Figure 8A shows a tube 110 as in Figure 7, with a section indicated which is shown enlarged in Figure 8B, which in turn shows the section of that panel enlarged in Figure 8C. As the movement upward of the microbanding tube causes the capillary tube to move toward the tube bottom, the difference in pumping rates of the two pistons attached to the flow cell causes fluid to flow up the capillary where it is diluted as described by the combined action of pumps 122 and 124 of Figure 7. However, the amount of fluid drawn into the capillary 117 is much greater than the volume of fluid effectively displaced from the banding gradient by the relative movements of the capillary and the microbanding tube. This volume is replaced by fluid flowing into tube 115 through cap 116 which is initially allowed to flow in until the tube 110 is full. This fluid is much less dense than the density of the fluid at the top of the gradient in the microbanding region, and causes minimal disturbance in the gradient. As shown in Figure 8B, the capillary 117 slowly approaches virus band 144, and, as shown in Figure 8C, a small amount of gradient liquid 145 is diluted by a larger amount of supernatant fluid 114 as it flows up the capillary. In this manner, a sharp band of virus particles 144 is diluted and moves through the flow cell as, volumetrically, a larger band, but with little effective loss of resolution. This technique provides the dilution necessary to make counting of individual virus particles feasible and accurate. The amount of dilution can be controlled such that the concentration of microorganisms in the capillary tube is less than one-half or one-tenth, or one-hundredth, or one-thousandth, or one ten-thousandth, or one-millionth, or one-billionth of the concentration in the band in the lower region of said tube.

In addition to counting the particles or determining the titre of a virus, the amount of DNA in the virus or other microbe can be determined for individual particles. In this aspect of the invention, the amount of DNA in the particles is measured by flow fluorescence analysis (Goodwin et al., 1993) or epifluorescence analysis (Jing et al., 1998). In this manner, yeast, bacteria, mycoplasma and virus can be distinguished as groups. For example, it is known that viruses contain $5\text{--}200 \times 10^3$ bases or base pairs, *E. coli*, a typical bacterium, contains 4×10^6 base pairs, while a typical yeast cell contains 1.3×10^7 base pairs. Thus, an estimate of the amount of DNA or RNA present allows the class of an infectious agent to be determined.

Thus, the size of a genome can be determined. In this embodiment of the invention, the genome is extracted from the microorganism band and immobilized on a solid support. The

immobilized DNA is stained and electronically imaged using an epifluorescence microscope (Jing et al., 1998). The length of the individual nucleic acid molecules can then be measured.

The technique of microbanding is useful not merely for staining the virus with dyes and being able to count the virus particles. Once the viruses from a biological sample have been highly purified and concentrated by the two dimensional centrifugation technique as described above using microbanding centrifuge tubes, the viruses are amenable for use in many other assays.

When an infectious agent is banded in a microbanding tube, the band may also be judiciously removed using a capillary needle in a volume of a few microliters, diluted to 5 mL or more with very dilute buffer or deionized water to dilute the gradient materials by a factor as large as 1,000, and then pelleted in a fresh microbanding tube. The supernatant may then be carefully withdrawn by a suction capillary, and the virus or other agent resuspended in approximately 1 microliter using a syringe made, for example, of fine Teflon® tubing fitted with a very small stainless steel wire plunger to fit. The sample may then be transferred to a mass spectrometer target, mixed with a matrix dye, and used for matrix assisted laser desorption ionization time of flight mass spectrometry (MALDI-TOF-MS) to determine directly the masses of viral coat proteins or of bacterial cell proteins. Technology described for sample concentration may also be applied, without a matrix dye, to electrospray or other mass spectrometric analysis systems, including the detection of intact viral mass.

A system similar to that shown in Figure 7 may also be used to produce the equivalent of molecular restriction fragment length maps of DNA molecules using restriction enzymes. For this work, virus or bacterial particle bands may be diluted and sedimented as described, after which the DNA may be extracted using detergents or other reagents well known in the art, treated with a restriction enzyme and a fluorescent dye, and the fragment sizes determined by flow cytometry (Goodwin et al., 1993; Hammond et al., U.S. Patent No. 5,558,998). Extracted DNA may also be immobilized on a solid support, stained with a fluorescent dye, and photographed using an epifluorescence microscope to determine the length of DNA molecules. The preparation may then be treated with a restriction endonuclease, and the number and lengths of the oligonucleotide fragments determined (Jing et al., 1998). These data are then compared with a database listing the expected fragment lengths for different viral or bacterial species to identify each agent. DNA fragment lengths may also be determined by gel electrophoresis.

Fluorescence labeled antibodies may also be added to the particle suspension studied, and the presence or absence of the label in isopycnicly banded particles determined. This approach

is useful for specific identifications, and the use of a set of antibodies labeled with dyes having different and unique spectral characteristics allows the presence or absence of a series of agents to be determined. Alternatively antibodies labeled with chelators for rare earth's such as Europium and Terbium may be employed, in which case delayed fluorescence is measured.

5 Serum or plasma typically has a physical density between 1.026 and 1.031. Viruses typically have banding densities between 1.17 and 1.55 in cesium chloride, and at much lower densities in iodinated gradient materials such as Iodixanol or sucrose (Graham et al., 1994). The intermediate wash and reagent layers between the sample and the banding gradient must therefore have densities less than the density of the lightest virus to be banded. Buffers used to dissolve gradient material
10 for virus isolation include 0.05 M sodium borate, and 0.02 M sodium cyanide, both of which prevent bacterial growth.

With human serum or plasma, centrifugation sufficient to remove platelets and other particles having sedimentation coefficients of approximately 10^4 S is used before banding of virus particles.

15 The banding density of virus particles depends on the nucleic acid/protein ratio, and the presence or absence of lipids and lipoproteins. Hence attachment of specific identifying antibodies labeled with fluorescent dyes should not only allow identification by fluorescence but by a banding density change.

To assist in identifying particles by density, fluorescent particles of known density may be
20 included in the sample as shown in Figure 6. These particles may include known fixed and fluorescently stained virus or bacterial particles of known banding density, or very small fluorescently labeled or non-fluorescently labeled plastic beads. When polystyrene latex particles are coated with antibodies, their banding densities are increased appreciably, and the density may be further increased by reaction of the antibody-coated particles with the antigen for which the
25 antibodies are specific (Anderson and Breillatt, 1971). Antibody-coated fluorescent polystyrene beads may therefore be used not only to locate virus particles but to identify them.

The stains which are currently most useful are described in the Handbook of Fluorescent Probes and Research Chemicals, R.P. Haugland, ed., Molecular Probes, Inc., Eugene, Oregon (1996), which lists the abbreviated dye names, their chemical names, absorption and emission
30 maxima, and the filter combinations most used.

For work with human pathogens, safe operation and containment are important (Cho et al., 1966). Use of swinging bucket rotors, while optimal from a physico-chemical point of view, require

extensive manipulation and have more parts than an angle-head rotor. The tubes described in Figure 3E are designed to be used in angle-head rotors, and allow sedimenting particles to travel along a wall at one unchanging angle. Such rotors are easier to use and handle in containment than are swinging bucket rotors, however, sedimentation in an angle head rotor is far from ideal. Hence, the development of methods and devices for safely working with swinging bucket rotors is important.

High speed centrifuge tubes are notoriously difficult to seal effectively and are a potential source of infection to laboratory personnel. In practice, nearly all high speed swinging bucket rotor tubes are not themselves sealed, but are enclosed in a metal bucket which is sealed with a metal cap which does not seal the tube. The centrifuge tubes are therefore open when loaded, moved to the centrifuge rotor, inserted, and removed. It is very difficult to decontaminate the outside of an open tube containing a density gradient without disturbing the gradient. In the present application it is desirable to be able to effectively seal the plastic tubes in such a manner that the outside surfaces can be cleaned with a suitable disinfectant before the tubes are inserted into centrifuge shields, and to be able to handle them safely until they are scanned.

Sealing is done, as shown in Figure 9A, by inserting an annular ring seal 151, having a physical density less than that of water, into tube 152. Ring 151 is slightly tapered so that it fits very tightly into tube 152, and has a center hole which can be plugged and unplugged. In one embodiment, the center hole is threaded to accept a short, plastic flat-head screw. Initially two gradient components, including a lighter solution 153 and a denser solution 155, are introduced to the bottom microbanding region with a small air bubble 154 between, as previously described. As shown in Figure 9B, the solution 156 containing the infectious agent or other particles is then introduced through tube 157, leaving bubble 158 to separate the sample from the upper gradient solution. The volume of sample introduced does not completely fill the centrifuge tube, leaving space 159 empty. The tube is then sealed, as shown in Figure 9C with, for example, a plastic flat head screw 160, leaving air bubble 160 in place. The outside of the tube is then sterilized by immersing it in a disinfectant such as a sodium hypochlorite or hydrogen peroxide solution, followed by a water wash and gentle drying – all with the tube in an upright position. After centrifugation, as shown in Figure 9D, the plastic upper seal has been driven downward by centrifugal force a small distance as the entrapped air rises around the plastic seal. However, since the seal has a density less than water, it is retained at the top of the liquid sample, leaving a small lip 160 which may be grasped with a hemostat to remove the tube from the centrifuge shield. During centrifugation, the infectious particles are sedimented out of liquid 163 and produce band

164 in the gradient. The screw in the ring seal is then removed, and, as shown in Figure 9E, the supernatant liquid 165, which may contain a fluorescent dye, is removed through tube 166, leaving meniscus 167. As shown in Figure 9F, a laser beam 168, entering the tube from above, is then aligned with the tube, causing the banded infectious agent to emit fluorescent light for detection as previously described.

When a step gradient containing various reagents in addition to those used for isopycnic banding is employed, as illustrated in Figure 2, the discs used to separate the several solutions rise to the top and would not allow the use of vertical laser illumination as shown in Figure 9F without removing the seal and the discs. In this instance, side illumination, as illustrated in Figure 6 would be employed.

The laser or delayed fluorescence systems can be completely contained, the mechanical operations done remotely through small stepping motors, and the tubes moved in and out of the contained system under remote control.

These techniques can be combined with mass spectrometry and fluorescence-based restriction fragment mapping to allow rapid diagnosis and identification of infectious agents. However, the estimates of the masses of individual proteins are generally taken from published sequence data, and do not include numerous posttranslational modifications. Mass spectrometric data bases must be created to include actual mass measurements of different microorganisms. In addition, virion protein mass measurements will allow the detection of many genetic variants. However, for many studies of microorganisms, including development of data bases, the key problem has been the development of methods for systematically providing highly concentrated and purified microsamples of microorganisms from patient samples, natural waters, and from tissue culture fluids. This problem is solved by the present invention.

An alternate embodiment of a virus detecting system, depicted in Figure 10, is configured to detect viruses without the use of a dye, such as the fluorescent dyes mentioned above with respect to the system depicted in Figure 5. The virus detecting system depicted in Figure 10 includes many of the components of the system in Figure 5, but instead of detecting fluorescence from a fluorescent dye, the system in Figure 10 measures scattered light emitted from the microbanded viruses.

The scanning and detection system illustrated schematically in Figure 10 includes a mount 61 that supports a microbanding tube 60 in a vertical position. The mount 61 is supported by goniometers 62 and 63 which are in turn supported by X-Y movements 64 and 65 in such a manner as to align and center the microbanding section of tube 60 with respect to a laser beam 66.

Specifically, the X-Y movements **64** and **65** are for orthogonal adjustments in a horizontal plane. The goniometers **62** and **63** are each adjustable about perpendicular conical surfaces that have center lines that pass through a portion of the tube **60**. By adjusting the positions of the goniometers **62** and **63** and X-Y movements **64** and **65** it is possible to align the particle zone **70** with the laser beam **66**.

5 The laser beam **66** is generated by a laser **67**, which may be an argon ion laser producing, for instance, coherent light at 458, 488, 496, 502, and 515 nm. One such laser producing device is the Omnicrome 5025 which is an argon laser. Some lasers include internal filters or other internal wavelength determining means such that only a selected wavelength of light is emitted from the laser. For such lasers, no external filter is necessary for producing coherent light at a single
10 frequency, although external filters may be used if desired or if necessary for using a different laser light source. For the configuration of the present invention depicted in Figure 10, polarized light is desirable and therefore the laser beam **66** may alternatively be provided by any laser light source that produces polarized light at a single frequency. The inventor has determined that smaller wavelengths are more advantageous in the light scattering measurement of the present invention. Specifically, a frequency of polarized laser light at 488 nm produces optimal results, as is explained
15 in greater detail below.

 The laser beam **66** is reflected down into the microbanding tube **60** by dichroic mirror **69**. A banded particle zone **70** is photographed or electronically scanned by camera **71** without the need of a filter. The entire system may be enclosed to eliminate stray light. A camera **71** focused on the
20 banded particle zone **70** measures the light scattered therefrom due to the laser beam **66** trained on the particle zone **70**.

 Planar polarized light, such as the laser light produced by the laser **67**, includes a magnetic component **H** and an electric component **E** that are perpendicular to one another. Specifically, for planar polarized light, the electric component **E** travels in a plane that is perpendicular to the
25 magnetic component **H**. The orientation of the laser beam **66** is such that as the laser beam **66** passes through the banded particle zone **70**, the plane defined by the electric component **E** is normal (perpendicular) to the line of sight from the camera **71**, as is shown in Figure 15. Note that Figure 15 does not include the tube **60** in order to provide greater clarity, but does show the banded zone **70**.

30 The inventor has discovered that with the laser beam in the above described orientation, shown in Figure 15, greater sensitivity is achieved in the measurement of scattered light emitting

from the banded zone 70 compared to measurement of fluorescence emitted from dyes that may be in the banded zone 70.

The present invention is described by reference to the following Examples, which are offered by way of illustration and are not intended to limit the invention in any manner. Standard techniques well known in the art or the techniques specifically described below were utilized.

Example 1

To illustrate the use of microbanding tubes, experimental studies have been carried out using a small single-stranded non-pathogenic DNA virus ϕ X 174. Approximately 10^{10} virus particles which had been purified by isopycnic banding in CsCl in a microbanding tube such as shown in Figure 2A were suspended in 5 mL of 0.05 M borate buffer, were pelleted in a microbanding tube at 35,000 rpm in a swinging bucket rotor, and were resuspended in approximately 3 μ L for analysis. The analyses were done on a PerSeptive Biosystems DESTRA instrument with an extraction delay time set at 150 nseconds using a matrix of sinapinic acid. Bovine insulin (MW=5,734.59) and horse heart apomyoglobin (MW=16,952) were used as 1 and 2 pmole standards. The results are shown in Table 1. The ϕ X 174 masses for virion capsid proteins F, G, H and J are calculated from published sequence data. The differences between the calculated and experimental values for F and H are probably due to posttranslational modifications. The probability that an unrelated virus could have subunits of the same masses listed is vanishingly small. However, even more definitive protein identifications can be made by treating viral proteins with proteolytic enzymes such as trypsin and determining the masses of the peptide fragments produced. Computer programs are available which calculate the sizes of fragments of proteins of known sequence by well characterized enzymes. Such programs include Protein Prospector (available from the University of California, San Francisco) and ProFound (available from Rockefeller University).

Table 1

Mass Spectrometric Analysis of ϕ X 174 Virion Proteins

<u>Protein</u>	<u>Calculated Mass</u>	<u>Experimental Mass</u>	<u>Mass Difference</u>	<u>% Difference</u>
F	48,351.53	48,407.4	+55.9	0.12 %
G	19,046.73	19,046.7	0.0	0.00 %
H	34,419.25	34,466.1	+46.9	0.14 %
J	4,095.78	4,097.03	+1.2	0.03%

This example demonstrates that highly purified and concentrated suspensions of microorganisms can be isolated from biological samples such as, but not limited to, patient samples such as plasma, urine, feces and tissues, natural water and tissue culture fluids. This example further demonstrates that such purified and concentrated microorganisms can then be identified, for example, using mass spectrometry to identify viruses.

Example 2

The inventor conducted an experiment to demonstrate the greater sensitivity realized by measuring scattered light with the camera 71 focused on a zone 70 with the line of sight of the camera 71 being normal to the plane of the electric component E of the laser beam 66. Specifically, lambda phage was purified by banding in cesium chloride and diluted to a titer of 1.5×10^{10} plaque forming units per mL. 1.5 mL of this preparation was mixed with 25 μ L of 1 mM YOYO-1 fluorescent DNA stain (marketed by Molecular Probes, IN), and left at refrigerator temperature for three days.

Parallel sets of microbanding tubes were set up with a step gradient set up in the microbanding region using 17 μ L of room temperature saturated cesium chloride at the bottom, followed by 17 μ L of a 1:10 dilution of the saturated solution in phosphate buffered saline (PBS), pH 7.2. This in turn was followed by 1.5 mL of 1:20 dilution of the same saturated solution in PBS. These solutions were then spaced with small air bubbles, a process made feasible by the very small diameter of the microbanding region of the tubes 60. Two parallel sets of tubes 60 were then filled, one set with PBS containing 3×10^8 stained virions, and the other set with PBS containing 3×10^8 unstained virions. All of the tubes were then centrifuged for 3 hours at 40,000 rpm in an SW41-Ti rotor in a Beckman L-70K ultracentrifuge. The tubes were vertically illuminated with polarized laser light at 488 nm using an argon ion laser. Images were captured using an Apogee CCD camera. Scatter was measured with both the dyed and non-dyed virus using 488 nm light, while the light emitted by the stained virus was measured using 510 nm light using an interference filter.

Figure 12 shows the scatter image generated by the camera 71 of one microbanding zone 70, while Figure 14 shows the fluorescence image generated by the camera 71 of the same microbanding zone 70. Figure 11 shows a computer generated graph of the light intensity detected by the camera 71 based upon the image in Figure 12, and Figure 13 shows a computer generated graph of the fluorescent light intensity based upon the image in Figure 14. The intensity units (the numbers on the vertical axis) on the graphs in Figures 11 and 13 are the same. However, the

depicted graphs were automatically scaled by the computer to fit a predetermined portion of the computer monitor. As can be seen by comparing the graphs in Figures 11 and 13, the peak height of the line scan for scatter was approximately 2300 arbitrary intensity units, while the peak height for the fluorescent virus was only 540 arbitrary intensity units. Clearly, detection of scatter is more sensitive than detection fluorescence under the conditions employed.

An advantage of light scatter measurement is that it is possible to determine the particle size by measuring the angle of maximum light scatter. Therefore, more information is obtainable from measurement of light scatter than from measurement of fluorescence. Further, some fluorescent dyes degrade after exposure to laser light. Light scatter is not affected by duration of exposure to laser light.

The inventor has further determined that the shorter the wavelength of light from the laser 67, the greater the amount of scattered light resulting in an increase in sensitivity. Specifically, several of the frequencies of polarized light produced by the laser 67 were used, one frequency at a time, in order to measure light scatter from a single microbanding zone 70. A polarized beam of light of 488 nm yielded the best results.

It is also possible to measure both fluorescence and light scatter on the zone 70 by using the camera 71 and moving at least filter 72 into position as indicated by the arrows in Figure 16. Note that the filter 68 is depicted, but is only necessary if the laser 67 outputs several wavelengths of light. The line of sight of the camera 71 is normal (perpendicular) to the plane with the electric component E. The camera 71 is first used to measure quantitatively the light intensity of light scatter from the zone 70 without the filter 72. The filter 68 is required to allow only one frequency of light if the laser 67 outputs several frequencies of light. Thereafter, the filter 72 is moved into position in front of the camera 71 and fluorescence is measured to confirm that the particles are viruses and not, for instance, dust, dirt or other non-essential components.

It should be understood that fluorescence measurement is not dependent upon the orientation of polarized light because the fluorescence emitted by the dye is excited by the laser light to emit light and does not necessarily reflect the laser light.

By the above described method and apparatus it is possible to obtain information from a single zone 70 by measuring both light scatter and fluorescence.

Another aspect of the invention is the addition of a polymerizable material, such as acrylamide, in the lower portion of the tube 60 that is hardened after centrifugation such that the banded virus may be fixed in position within the tube 60 for later excising and analysis. For

instance, sodium alginate may be introduced into the tube **60** and, after centrifuging, calcium ions may be added to the surface causing the mixture in the tube **60** to gel. The semisolid mass in the tube can be removed, and the zone with the virus removed for further study.

The above described methods and apparatus can be used to detect or isolate viruses from plants, animals or environmental sources.

While the invention has been disclosed in this patent application by reference to the details of preferred embodiments of the invention, it is to be understood that the disclosure is intended in an illustrative rather than in a limiting sense, as it is contemplated that modifications will readily occur to those skilled in the art, within the spirit of the invention and the scope of the appended claims.

LIST OF REFERENCES

- Anderson, N.G., ed. (1966). "The Development of Zonal Centrifuges." *Natl. Cancer Inst. Monogr.* 21.
- Anderson, N.G. et al. (1966). "Separation of subcellular components and viruses by combined rate- and isopycnic-zonal centrifugation." In The Development of Zonal Centrifuges. N.G. Anderson, ed. *National Cancer Institute Monograph* 21: 253-284.
- Anderson, N.G. et al. (1967). "Isolation of viral particles from large fluid volumes." In Transmission of Viruses by the Water Route, G. Berg ed., Interscience Publishers, NY, pp. 75-88.
- Anderson, N.G. (1968). "Analytical techniques for cell fractions. VIII. Analytical differential centrifugation in angle-head rotors." *Anal. Biochem.* 23: 72-83.
- Anderson, N.G. and Breillatt, J.P. (1971). "Isopycnetric serology: A new technique based on buoyant density changes in latex beads." *Nature* 231: 112-114.
- Bergh, O. et al. (1989). "High abundance of viruses found in aquatic environments." *Nature* 340: 467-468.
- Borsheim, K.Y. et al. (1990). "Enumeration and biomass estimation of planktonic bacteria and viruses by transmission electron microscopy." *Applied Environ. Microbiol.* 56: 352-356.
- Chang, Y. et al. (1994). "Identification of herpes virus-like DNA sequences in AIDS-associated Kaposi's sarcoma." *Science* 265: 1865-1869.
- Cho, N. et al. (1966). "Problems in biocontainment." *Natl. Cancer Inst. Monogr.* 21: 485-502.
- Claydon, M.A. et al. (1996). "The rapid identification of intact microorganisms using mass spectrometry." *Nature Biotechnology* 14: 1584-1586.
- Fenselau, C. (ed.) (1994). "Mass spectrometry for the characterization of micro-organisms." *ACS Symposium Series*, Vol. 541, ACS, Washington DC.
- Fredricks, D.M. and Relman, D.A. (1996). "Sequence-based evidence of microbial disease causation: When Koch's postulates don't fit." *J. NIH Res.* 8: 39-44.
- Gao, S.-J. and Moore, P.S. (1996). "Molecular approaches to the identification of unculturable infectious agents." *Emerging Infectious Diseases* 2: 159-167.
- Goodwin, P.M. et al. (1993). "Rapid sizing of individual fluorescently stained DNA fragments by flow cytometry." *Nucl. Acids Res.* 21: 803-806.
- Graham, J. et al. (1994). "The preparation of subcellular organelles from mouse liver in self-generated gradients of Iodixanol." *Anal. Biochem.* 220: 367-373.
- Hara, S. et al. (1991). "Abundance of viruses in marine waters: Assessment by epifluorescence and transmission electron microscopy." *Applied Environ. Microbiol.* 57: 2731-1734.
- Harris, W.W. et al. (1966). "Unusual particles in human plasma from leukemia and lymphosarcoma." In The Development of Zonal Centrifuges. *National Cancer Institute Monograph* 21: 289-394.

- Haugland, R.P. (1996). Handbook of Fluorescent Probes and Research Chemicals, 6th ed. Molecular Probes Inc., Eugene OR, p. 679
- Hennes, K.P. and Suttle, C.A. (1995). "Direct counts of viruses in natural waters and laboratory cultures by epifluorescence microscopy." *Limnol. Oceanogr.* 40: 1050-1055.
- Hennes, K.P. et al. (1995). "Fluorescently labeled virus probes show that natural virus populations can control the structure of marine microbial communities." *Applied Environ. Microbiol.* 61: 3623-3627.
- Hirons, G.T. et al. (1994). "TOTO and YOYO: New very bright fluorochromes for DNA content analysis by flow cytometry." *Cytom.* 15: 129-140.
- Holland, R.D. et al. (1996). "Rapid identification of intact whole bacteria based on spectral patterns using matrix assisted laser desorption ionization with time of flight mass spectrometry." *Rapid Commun. Mass Spectrom.* 10: 1227-1232.
- Jing, J. et al. (1998). Automated high resolution optical mapping using arrayed, fluid-fixed DNA molecules. *Proc. Natl. Acad. Sci. USA* 95: 8046-51.
- Kovacs, E. (1962). Chemical dissection of mammalian cells with liberation of biologically intact viruses. *Z. Naturforschg.* 17: 234-240.
- Krishnanurthy, T. et al. (1996). "Detection of pathogenic and non-pathogenic bacteria by matrix assisted laser desorption ionization time of flight mass spectrometry." *Rapid Commun. Mass Spectrom.* 10: 883-888.
- Liang, X. et al. (1996). "Determination of bacterial protein profiles by matrix assisted laser desorption ionization mass spectrometry with high performance liquid chromatography." *Rapid Commun. Mass Spectrom.* 10: 1219-1226.
- Linnen, J. et al. (1996). "Molecular cloning and disease association of hepatitis G virus: a transfusion-transmissible agent." *Science* 271: 505-508.
- Mazzone, H.M. (1998). *CRC Handbook of Viruses*. CRC Press, Boca Raton, p. 206.
- National Cancer Institute Monograph 21, *The Development of Zonal Centrifuges and Ancillary Systems for Tissue Fractionation and Analysis*, U.S. Department of Health, Education and Welfare, Public Health Service, 1966
- Samad, A.H. et al. (1995). "Mapping the genome one molecule at a time - optical mapping." *Nature* 378: 516-517.
- Simons, J.N. et al. (1995). "Identification of two flavi-virus-like genomes in the GB hepatitis agent." *Proc. Natl. Acad. Sci. USA* 92: 3401-5.
- Siuzdak, G. (1998). Probing viruses with mass spectrometry. *J. Mass Spec.* 33: 203-211.
- Suttle, C.A. (1993). "Enumeration and isolation of viruses." In Handbook of Methods in Aquatic Microbial Ecology, Chapter 15. Kemp, P.F., Sherr, B.F., Sherr, E.B. and Cole, J.J., eds. Lewis Publishers, Boca Raton, Fla., pp 121-134.
- Suttle, C.A. and Chen, F. (1992). "Mechanisms and rates of decay of marine viruses in seawater." *Appl. Environ. Microbiol.* 58: 3721-3729.

- Wommack, K.E. et al. (1992). "Distribution of viruses in the Chesapeake Bay." *Appl. Environ. Microbiol.* 58: 2965-2970.
- Yolken, R.H. and Torrey, E.F. (1996). "Viruses, schizophrenia and bipolar disorder." *Clinical Microbiology Reviews* 8: 131-145.
- Zeuzem, S. et al. (1996). "Phylogenetic analysis of hepatitis C virus isolates and their correlation to viremia, liver function tests, and histology." *Hepatology* 24: 1003-1009.

Patents

- U.S. Patent No. 3,519,400
- U.S. Patent No. 4,106,907
- U.S. Patent No. 4,624,835
- U.S. Patent No. 4,861,477
- U.S. Patent No. 5,342,790
- U.S. Patent No. 5,422,018
- U.S. Patent No. 5,489,396.
- U.S. Patent No. 5,558,998.

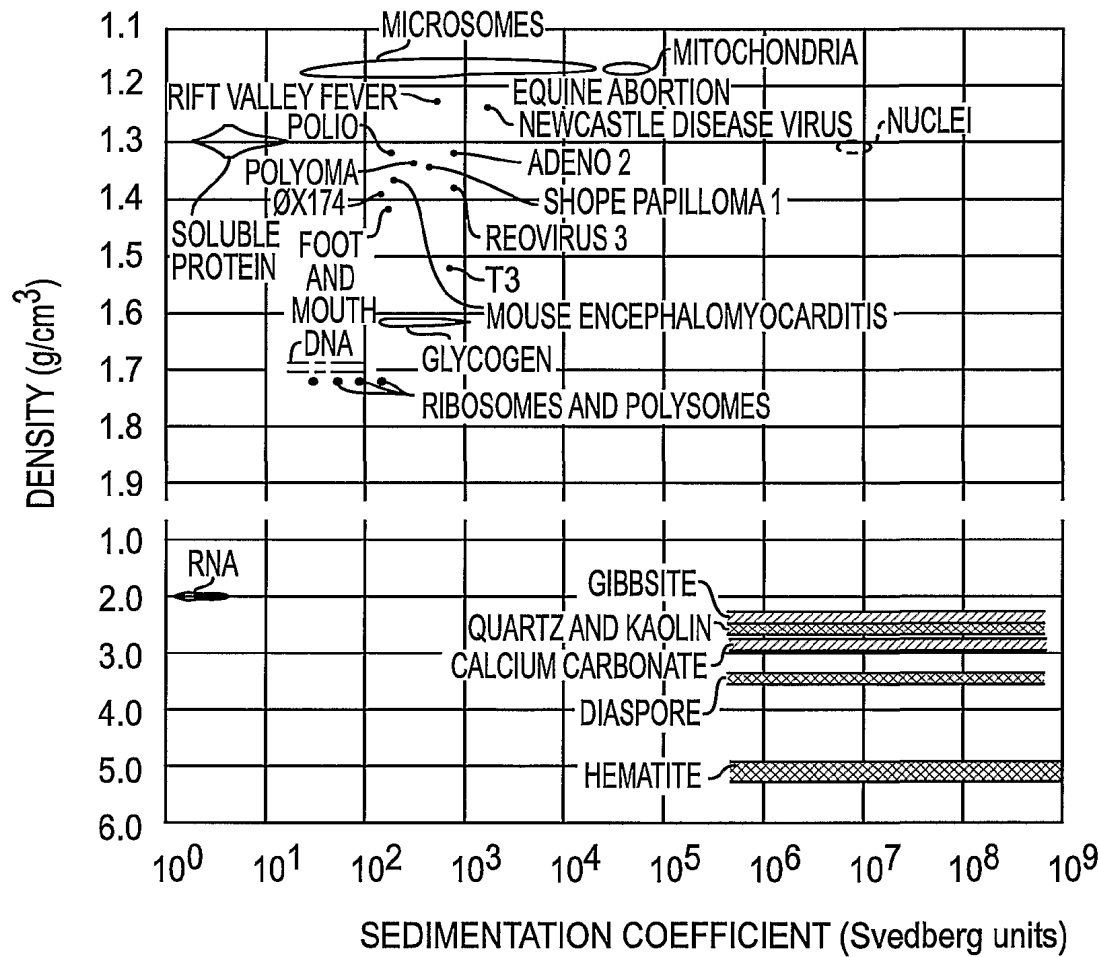
WHAT IS CLAIMED IS:

1. A system for measuring light emitted or scattered from a sample in a centrifuge tube wherein said system comprises:
 - a centrifuge tube holder to hold the centrifuge tube in a vertical position;
 - a light source which produces a beam of coherent polarized light; and
 - a detector which detects light from said sample, a line of sight of said detector being normal to a plane defined by the electric component of the coherent polarized light from said light source.
2. The system of claim 1, comprising in addition ,
 - goniometers onto which said centrifuge tube holder is mounted thereby enabling a centrifuge tube which is in said centrifuge tube holder to be oriented to match the vertical angle of the beam of coherent polarized light;
 - an X-Y movement to align the centrifuge tube with the beam of coherent polarized light in any X-Y direction; and
 - a mirror to deflect said beam of coherent polarized light through the centrifuge tube held in said centrifuge tube holder.
3. The system of claim 1, wherein said detector detects light scatter from the sample.
4. The system of claim 1, further comprising a computer having a monitor.
5. The system of claim 1, wherein said light source is a laser.
6. The system of claim 1, further comprising a filter disposed between the sample and the detector, said filter for allowing only fluoresced light to pass therethrough.
7. A method for measuring light emitted or scattered from a sample in a centrifuge tube wherein said method comprises the steps of:
 - centrifuging the centrifuge tube and sample to generate a microbanding zone thereby separating a portion of the sample;
 - passing a beam of coherent polarized light through the centrifuge tube;

detecting light emitted or scattered from said sample along a line of sight that is normal to a plane defined by the electric component of the coherent polarized light.

8. The method of claim 7, wherein scattered light from the sample is detected.
9. The method of claim 7, wherein the beam of coherent polarized light is generated by a laser.
10. The method of claim 7, further comprising the step of:
filtering the light from the sample thereby detecting only fluorescent light.
11. The method of claim 7, wherein both scattered light and fluoresced light are independently detected.

FIG. 1



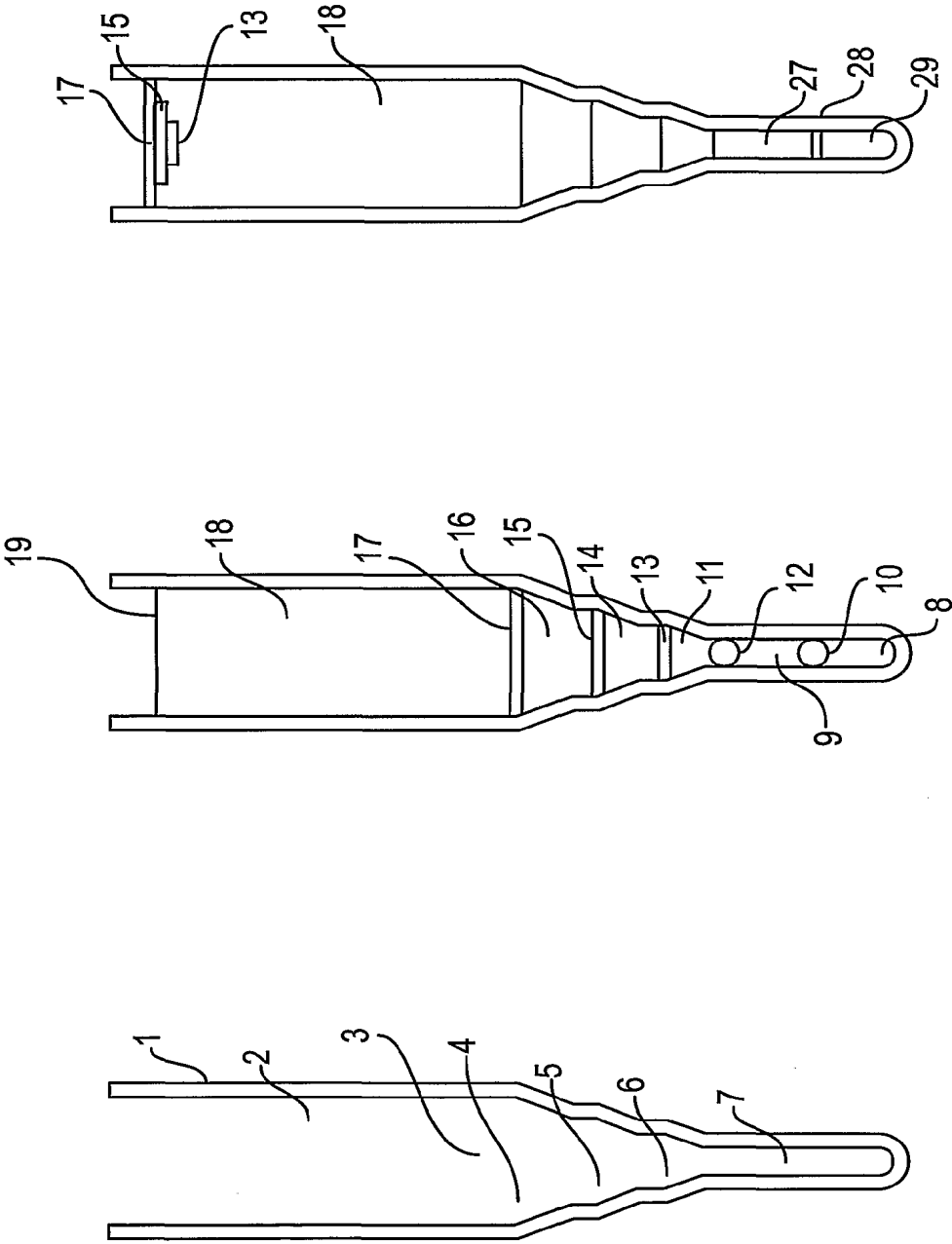


FIG. 2C

FIG. 2B

FIG. 2A

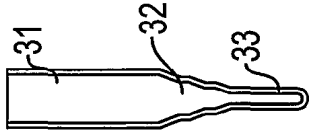


FIG. 3A

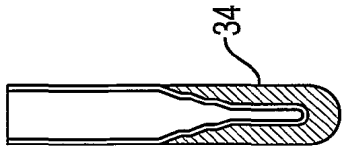


FIG. 3B

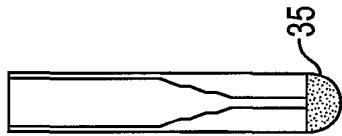


FIG. 3C

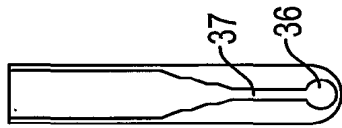


FIG. 3D

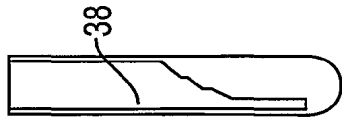


FIG. 3E

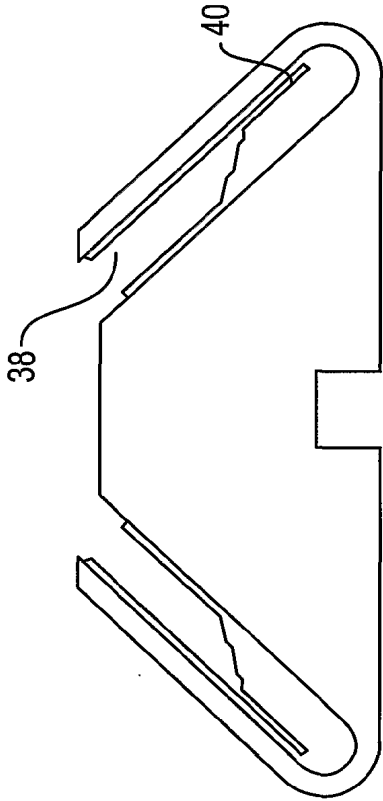


FIG. 3F

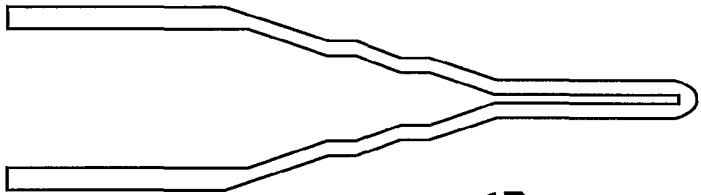


FIG. 3G

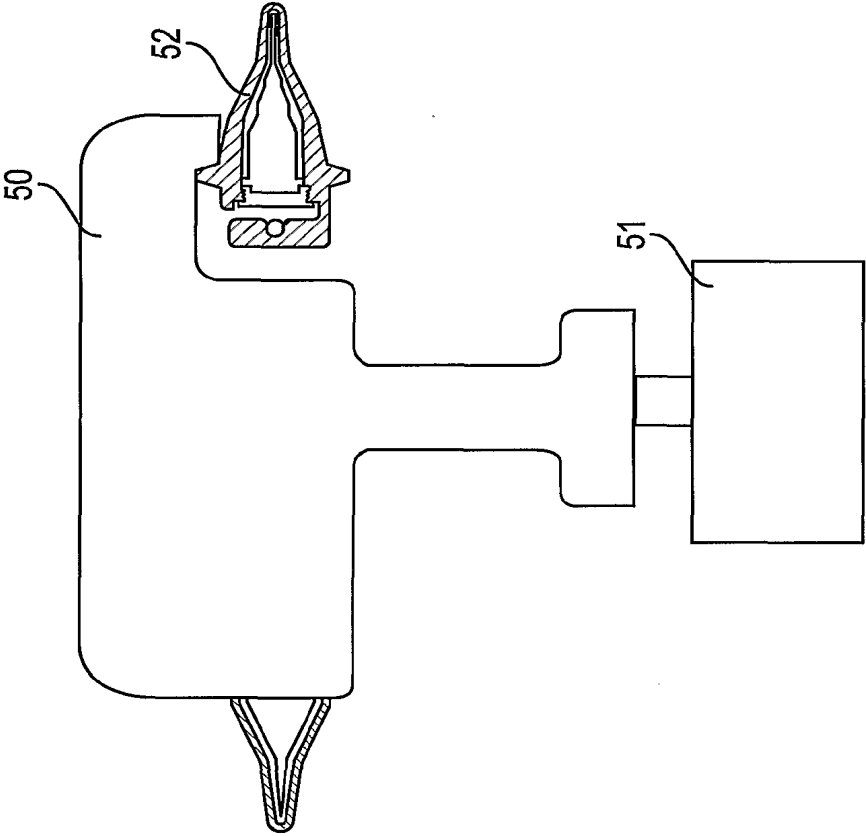


FIG. 4B

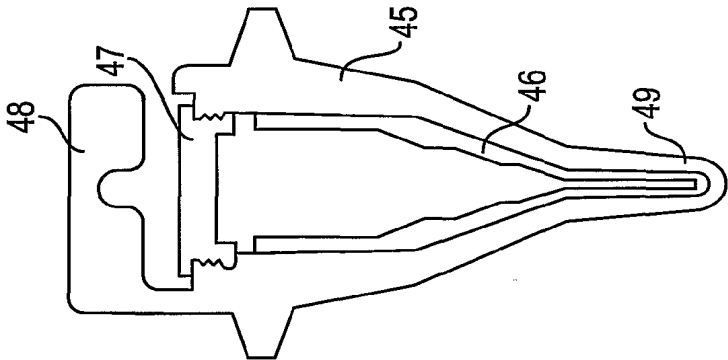


FIG. 4A

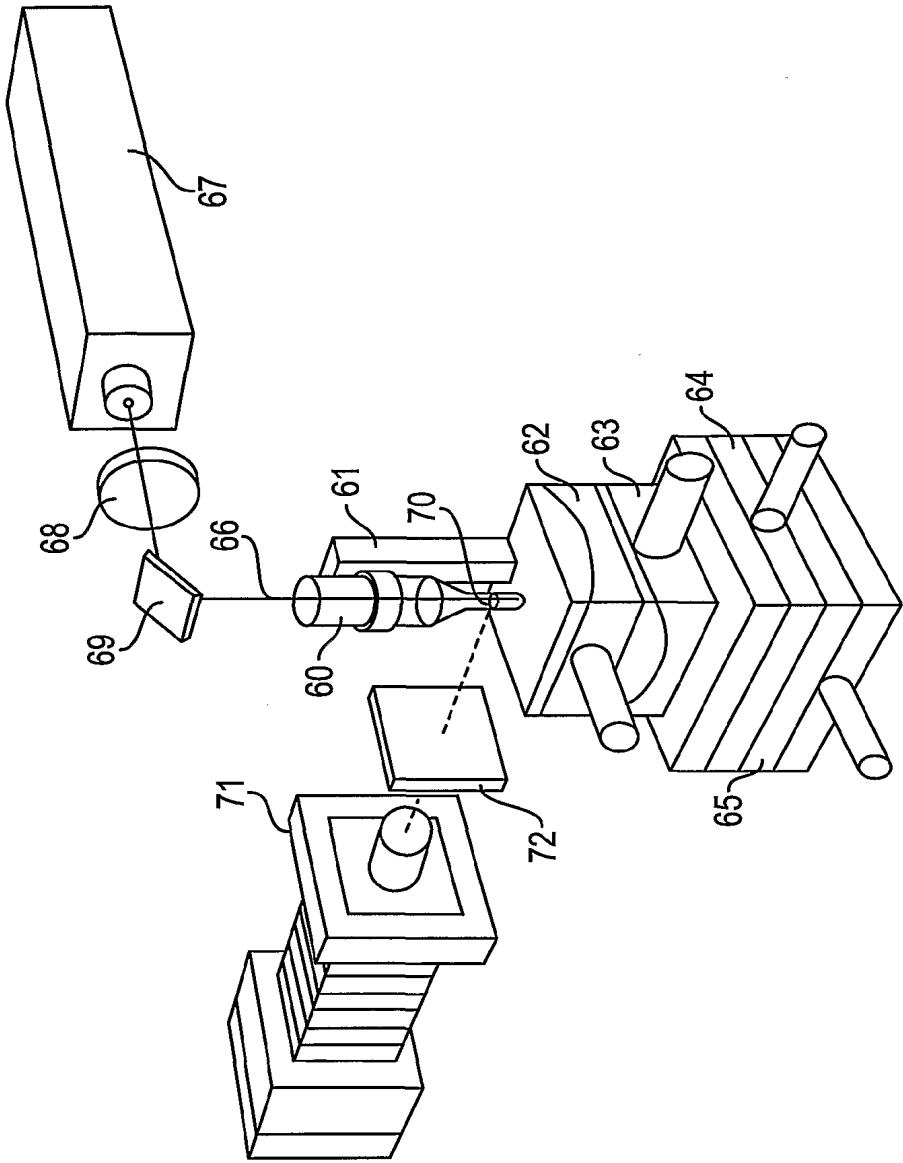


FIG. 5

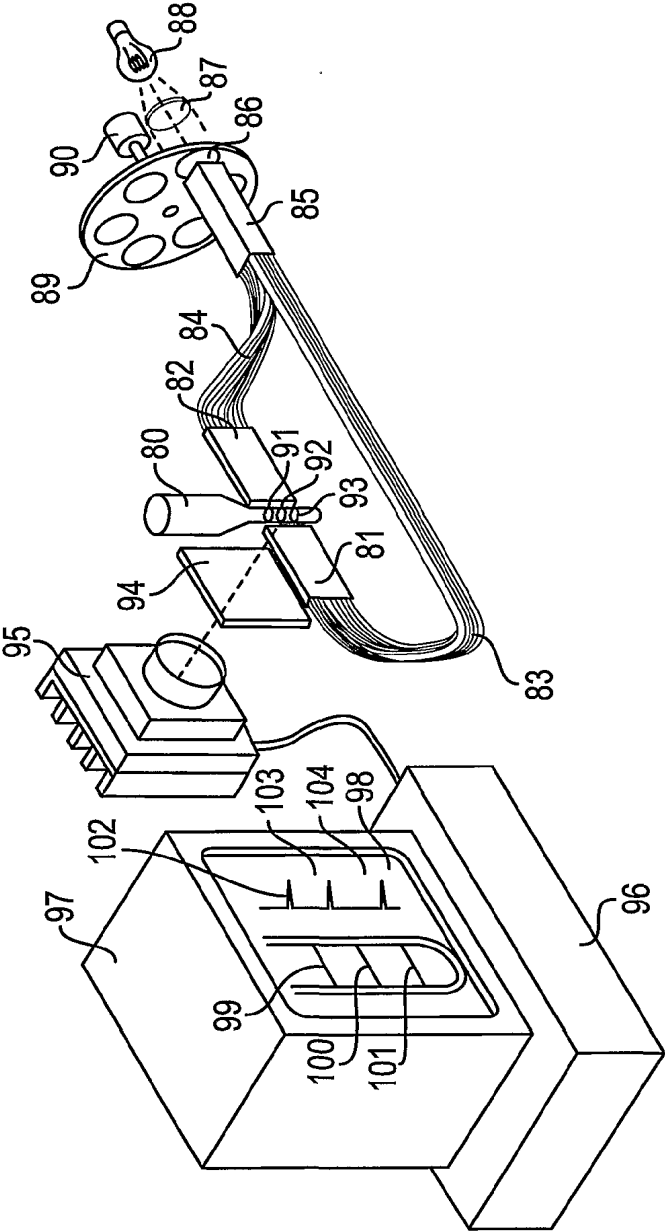


FIG. 6

7/16

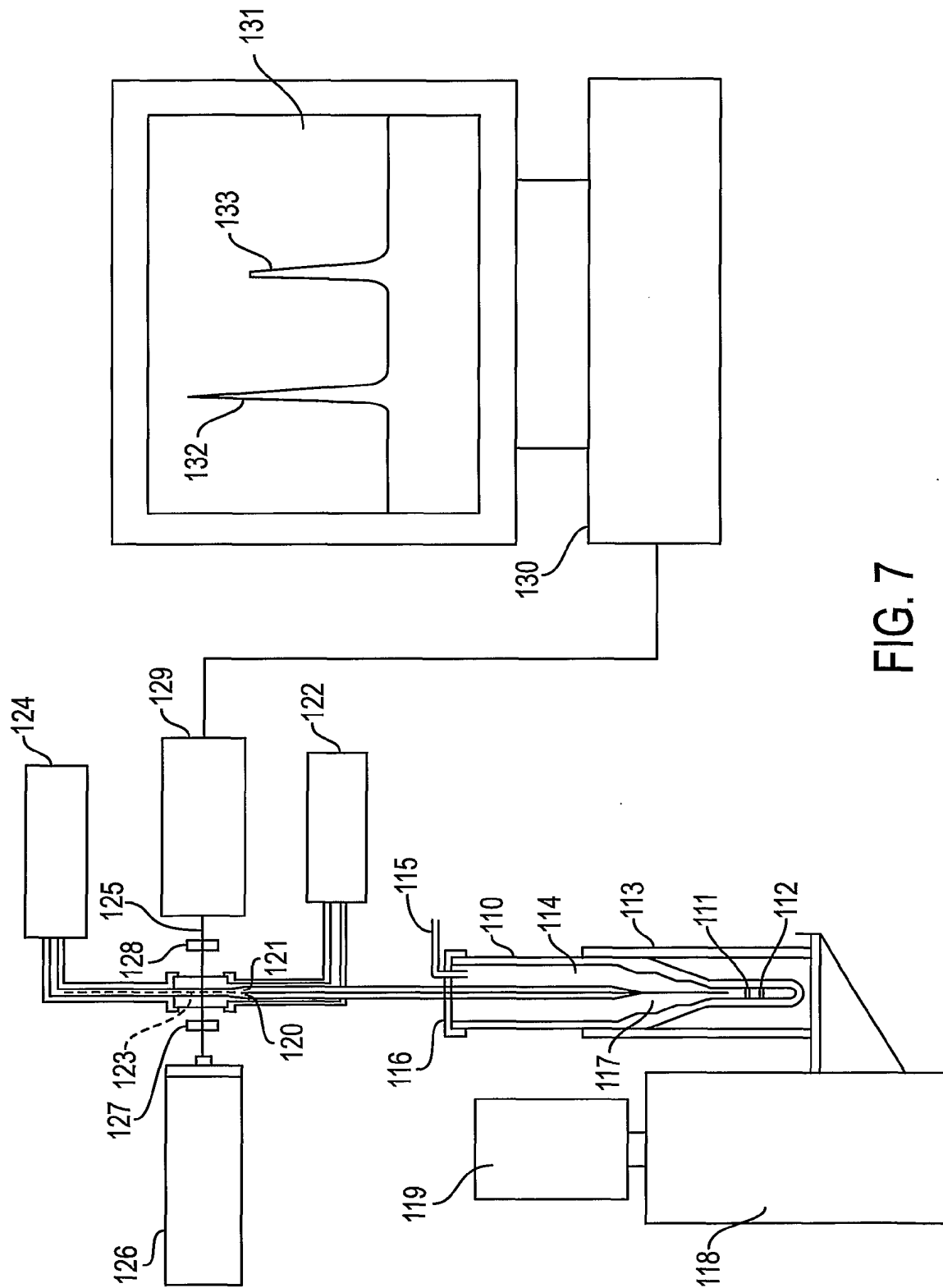


FIG. 7

8/16

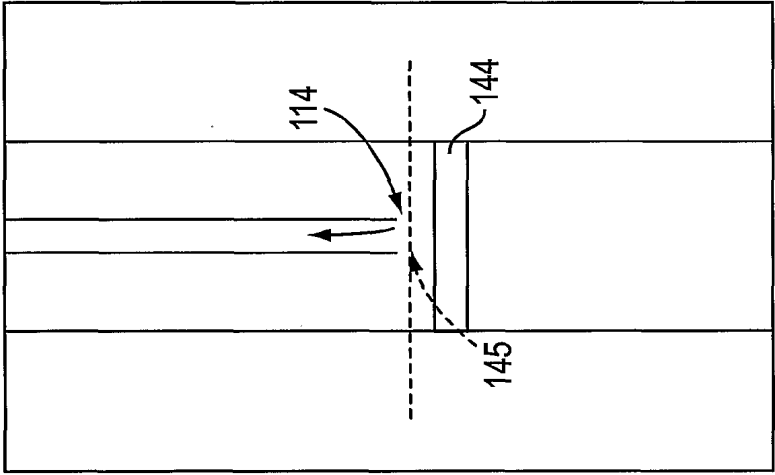


FIG. 8C

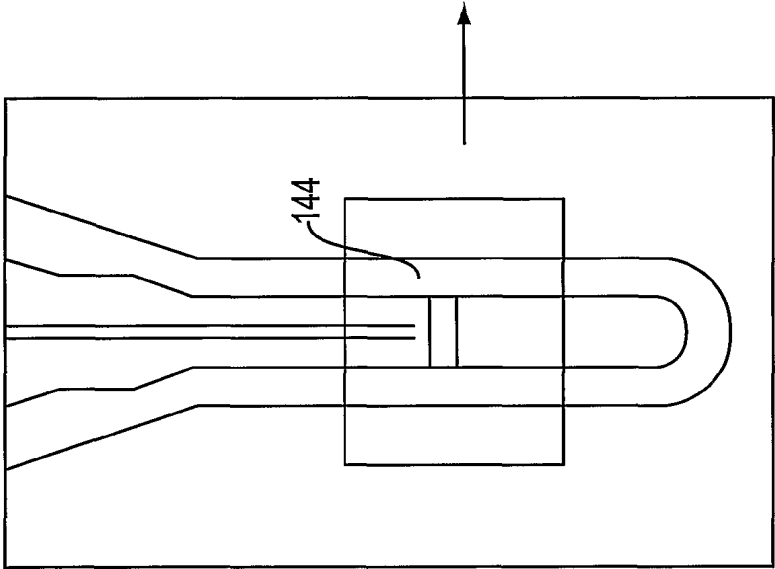


FIG. 8B

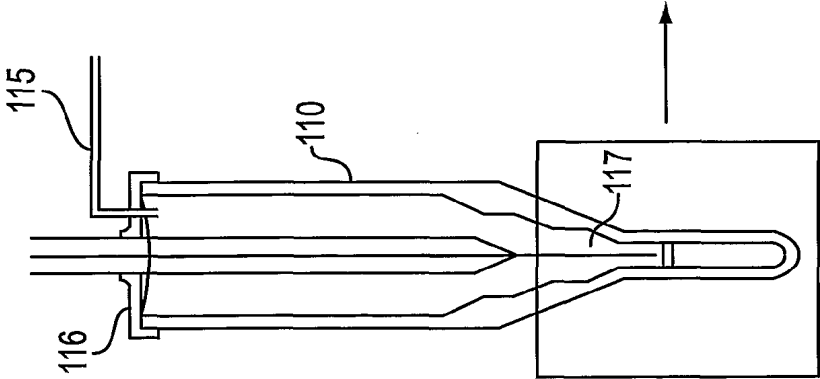


FIG. 8A

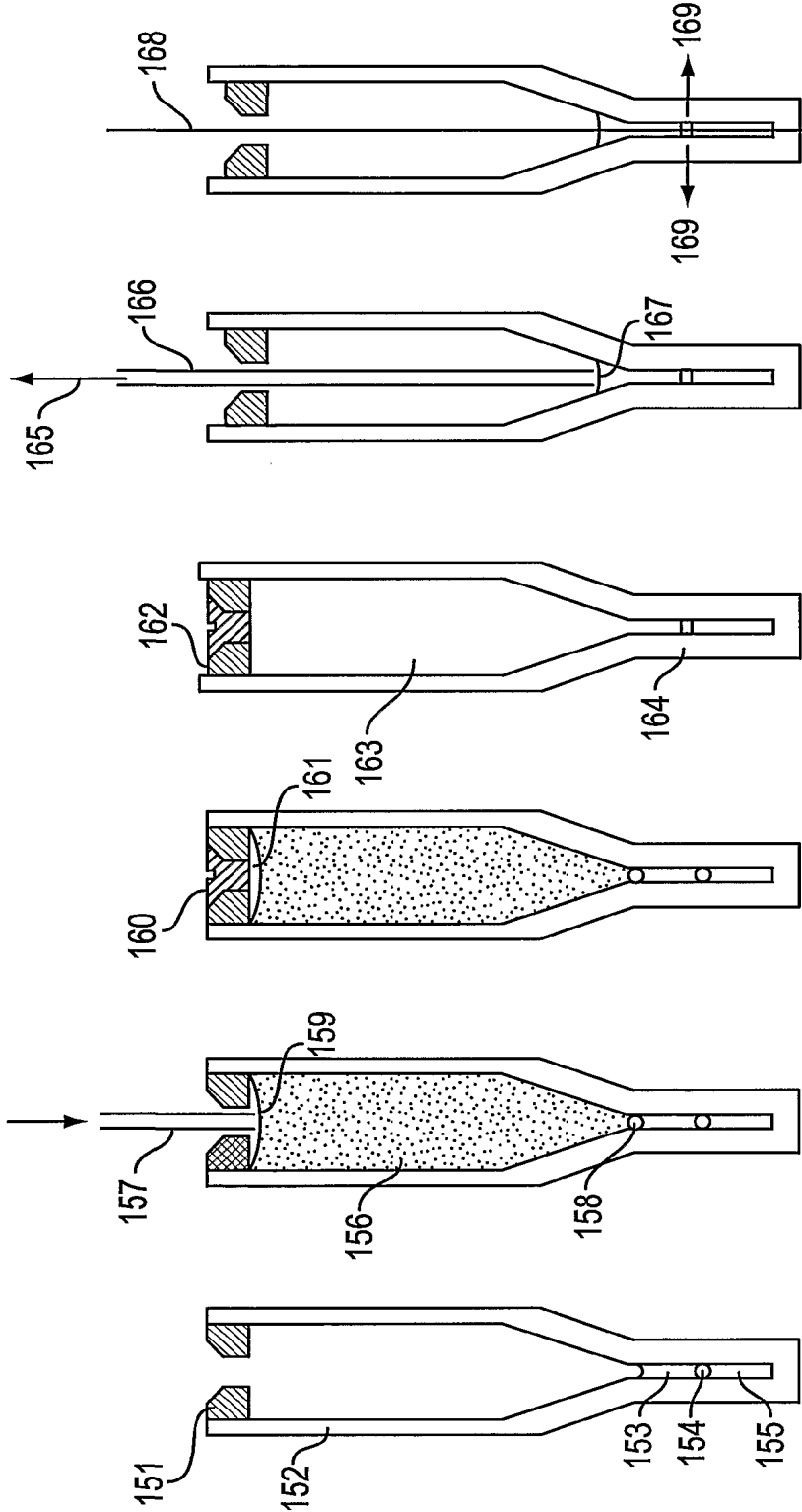


FIG. 9F

FIG. 9E

FIG. 9D

FIG. 9C

FIG. 9B

FIG. 9A

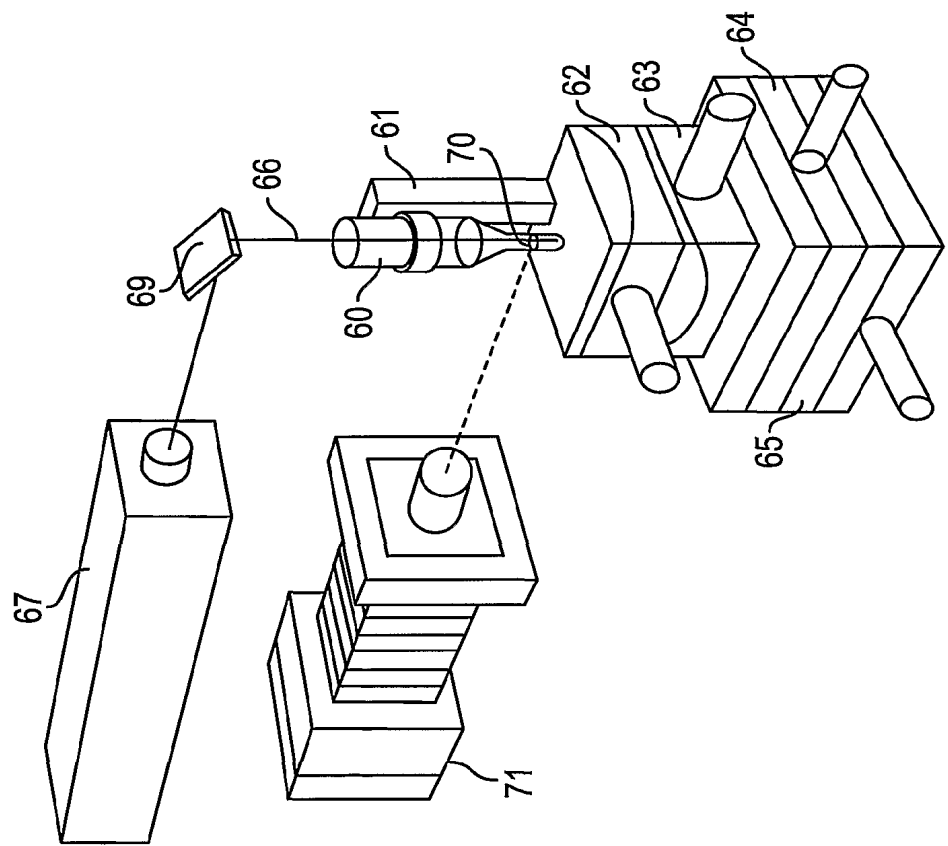


FIG. 10

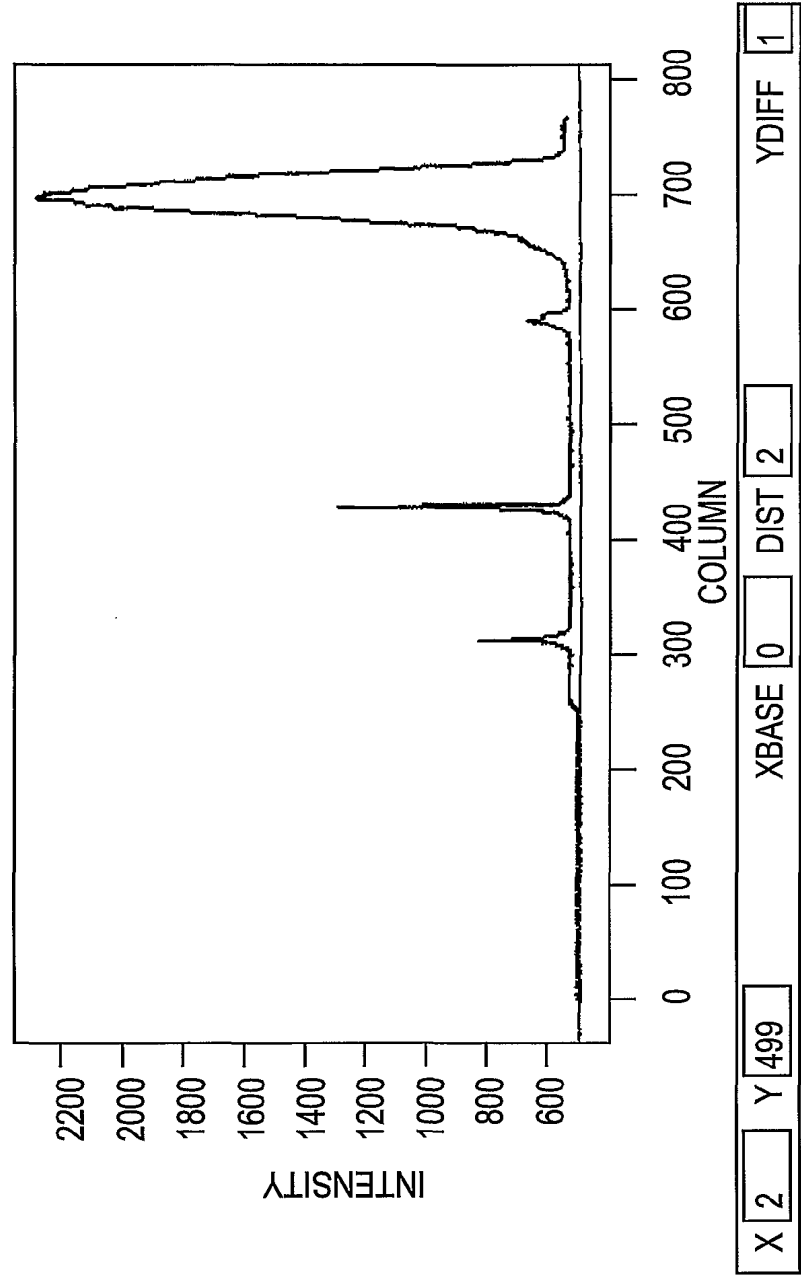


FIG. 11

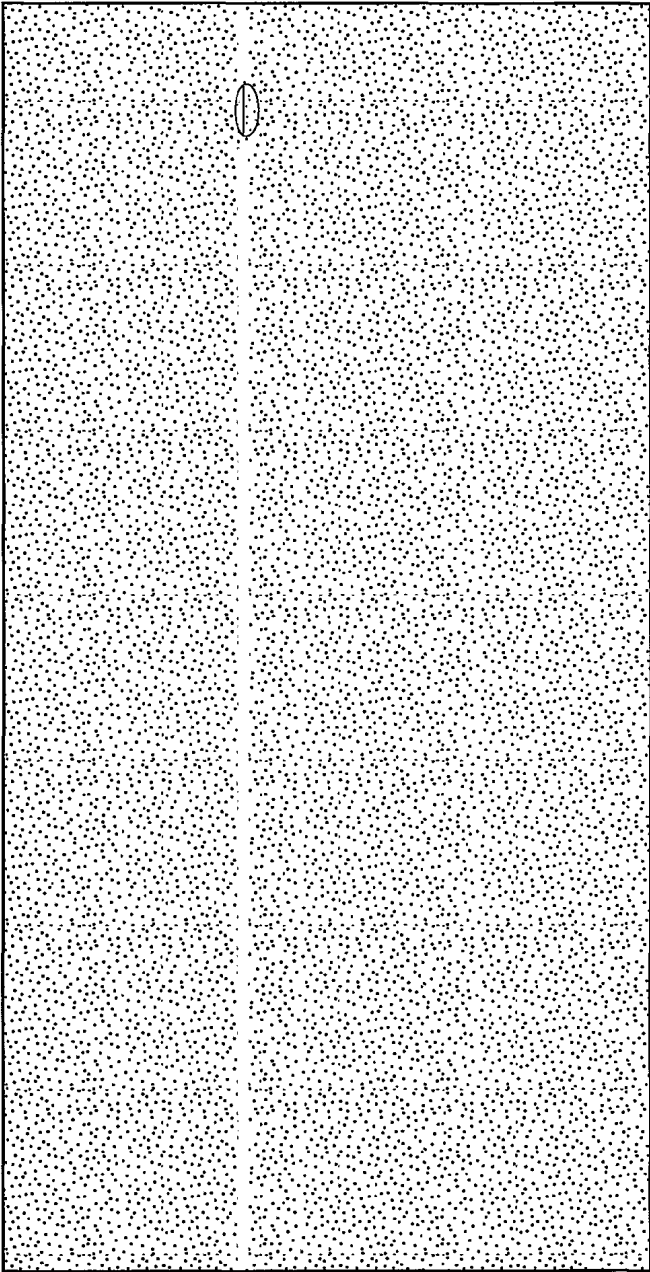


FIG. 12

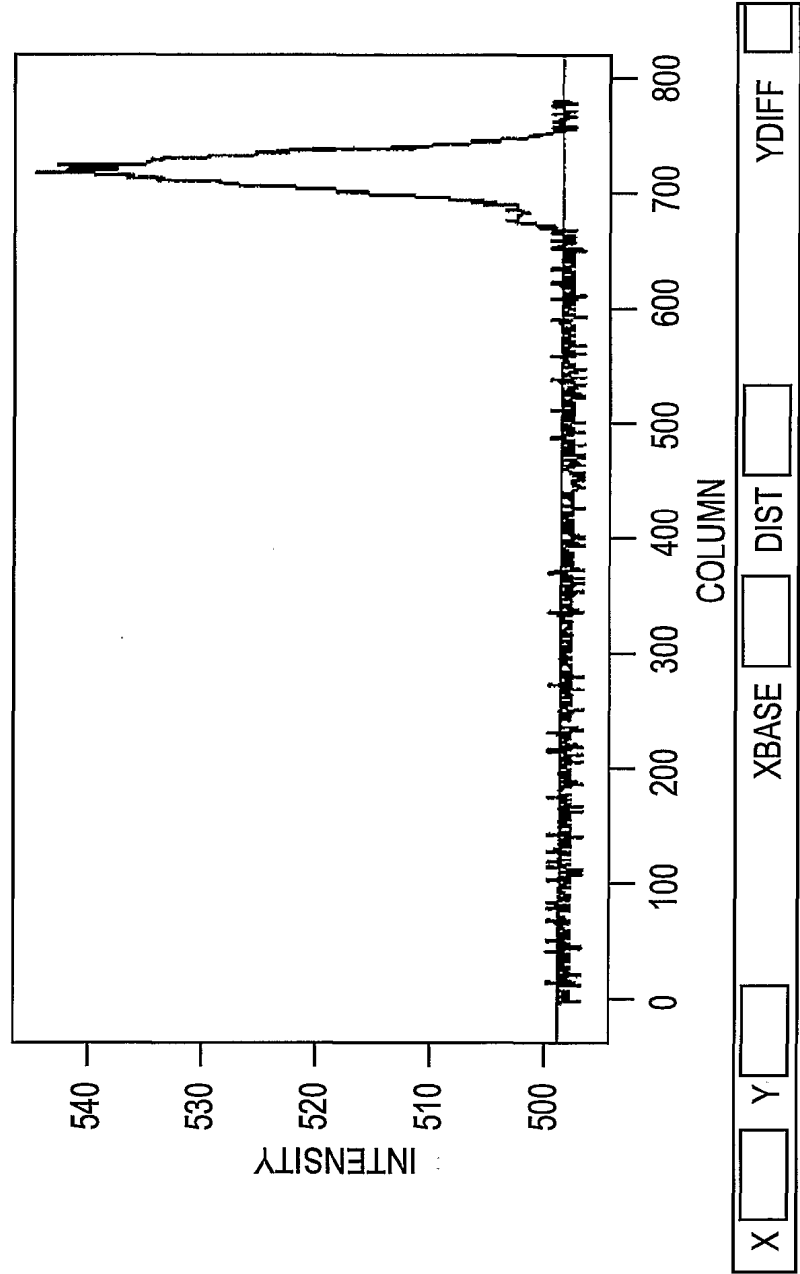


FIG. 13

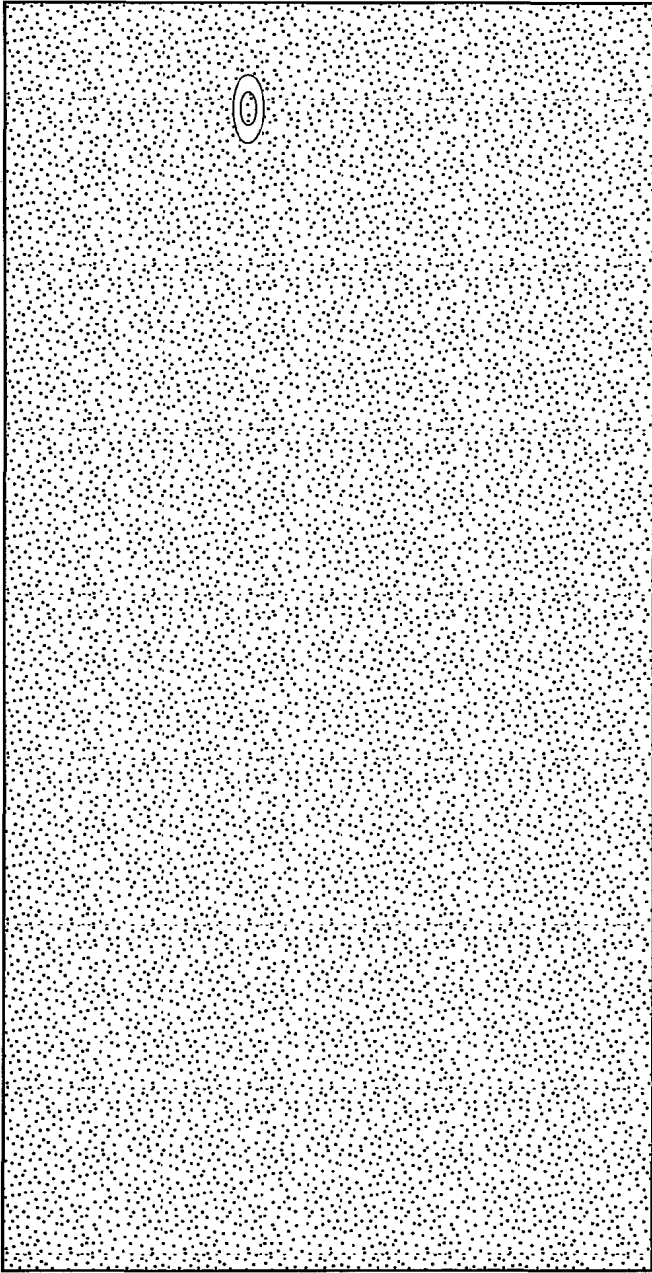


FIG. 14

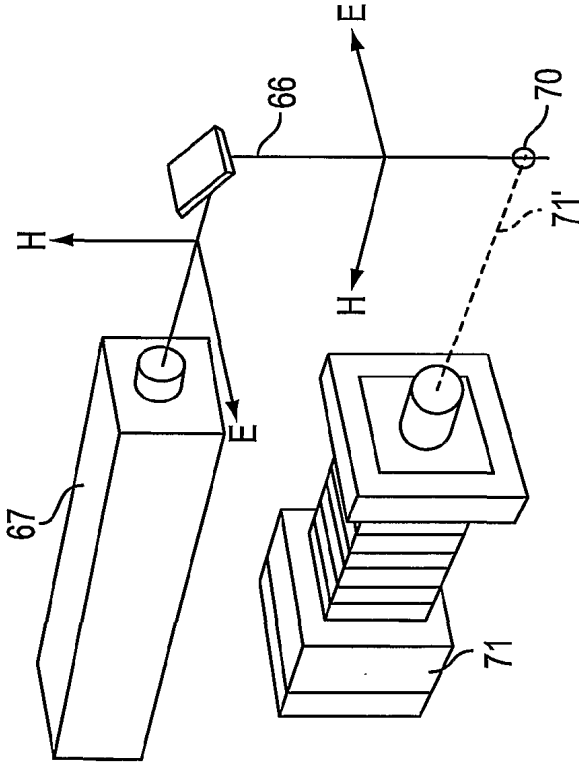


FIG. 15

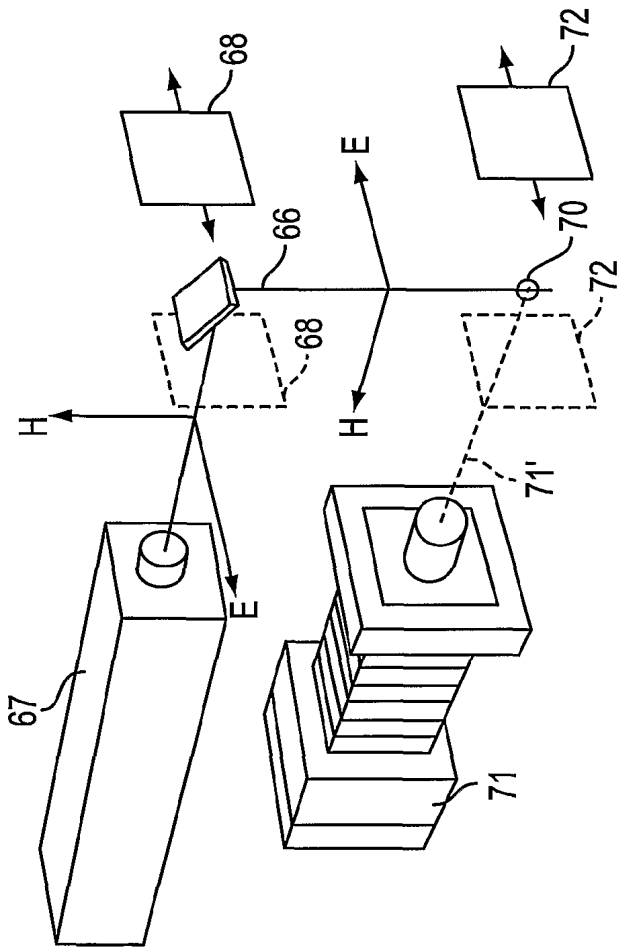


FIG. 16

INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 01/27529

A. CLASSIFICATION OF SUBJECT MATTER
 IPC 7 G01N21/51 G01N21/64

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 G01N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

WPI Data, PAJ, EPO-Internal

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 99 46047 A (ANDERSON NORMAN G ;ANDERSON N LEIGH (US); BIOSOURCE PROTEOMICS INC) 16 September 1999 (1999-09-16) page 10, line 6 - line 11 page 13, line 8 -page 14, line 16 claims 61-64; figures	1-10
Y	---	11
Y	WO 99 57533 A (CRANE DAVID OGILVIE ;CRANE ELECTRONICS (GB); EVERITT PETER WILLIAM) 11 November 1999 (1999-11-11) claim 7 -----	11

☐ Further documents are listed in the continuation of box C.



Patent family members are listed in annex.

* Special categories of cited documents:

- *A* document defining the general state of the art which is not considered to be of particular relevance
- *E* earlier document but published on or after the international filing date
- *L* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- *O* document referring to an oral disclosure, use, exhibition or other means
- *P* document published prior to the international filing date but later than the priority date claimed

- *T* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
- *X* document of particular relevance: the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
- *Y* document of particular relevance: the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.
- * & * document member of the same patent family

Date of the actual completion of the international search

19 March 2002

Date of mailing of the international search report

26/03/2002

Name and mailing address of the ISA

European Patent Office, P.B. 5818 Patentlaan 2
 NL - 2280 HV Rijswijk
 Tel. (+31-70) 340-2040, Tx. 31 651 epo nl.
 Fax: (+31-70) 340-3016

Authorized officer

Krametz, E

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/US 01/27529

Patent document cited in search report		Publication date	Patent family member(s)	Publication date
WO 9946047	A	16-09-1999	AU 3003099 A	27-09-1999
			CA 2322202 A1	16-09-1999
			EP 1062044 A2	27-12-2000
			WO 9946047 A2	16-09-1999
			US 6254834 B1	03-07-2001
			US 6340570 B1	22-01-2002
			US 6346421 B1	12-02-2002
<hr/>				
WO 9957533	A	11-11-1999	EP 1075646 A1	14-02-2001
			WO 9957533 A1	11-11-1999
<hr/>				